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**FLOW PREFERENCES OF  
AQUATIC INVERTEBRATES IN  
THE TONGARIRO RIVER**

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by

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# FLOW PREFERENCES OF AQUATIC INVERTEBRATES IN THE TONGARIRO RIVER

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## ABSTRACT

Aquatic invertebrates were collected from four sites along the Tongariro River in December 1990 to determine water depth and velocity preferences of the major invertebrate taxa, including five dominant Chironomidae (midge) taxa (three sites only). In addition, I investigated variations in flow preferences with size of nymphs of the mayfly genus *Deleatidium* spp. and of uncased caddisfly larvae belonging to the family Hydrobiosidae (most were *Hydrobiosis* and *Costachorema* species) at two sites to determine if flow preferences varied with larval size. Additional invertebrate sampling was carried out at a previously unsampled site in December 1992 to verify conclusions derived from the calculation of preference curves. It is intended that the resulting preference curves be linked with hydraulic surveys of the river and with information on the diet of blue duck and juvenile trout to determine the likely effects of different flow regimes on food supplies in the Tongariro River.

## 1. INTRODUCTION

The Tongariro River is the largest tributary of Lake Taupo. It receives drainage from the Kaimanawa Mountains in the east and volcanoes of Tongariro National Park in the west. The natural flow of the river has been substantially modified by a hydroelectric power scheme. Water is diverted from Moawhango River, Waihohonu Stream and upper tributaries of Whangaehu River into the Tongariro above Rangipo Dam. After passing through the turbines at Rangipo Power Station, the water re-enters the river above Poutu Intake which diverts much of it into Poutu Stream. Some of this water passes back into the Tongariro River via Poutu Stream (Fig. 1). Details of the power scheme, current flow management rules and fisheries issues are discussed in Stephens (1989).

The Tongariro River supports a nationally important trout fishery (Tierney 1988) and a population of the endangered blue duck (Cunningham 1991). Concern has been expressed about low juvenile trout production for several kilometres below Poutu Intake (Stephens 1989), and an apparent decline in the size of the blue duck population since the commissioning of the power scheme (Speedy and Keys 1992). These may be linked to decreased availability of food supplies brought about by a reduction in the area of suitable habitat for aquatic invertebrates, and/or a reduction in the quality of remaining habitat. As part of information gathering for the Tongariro

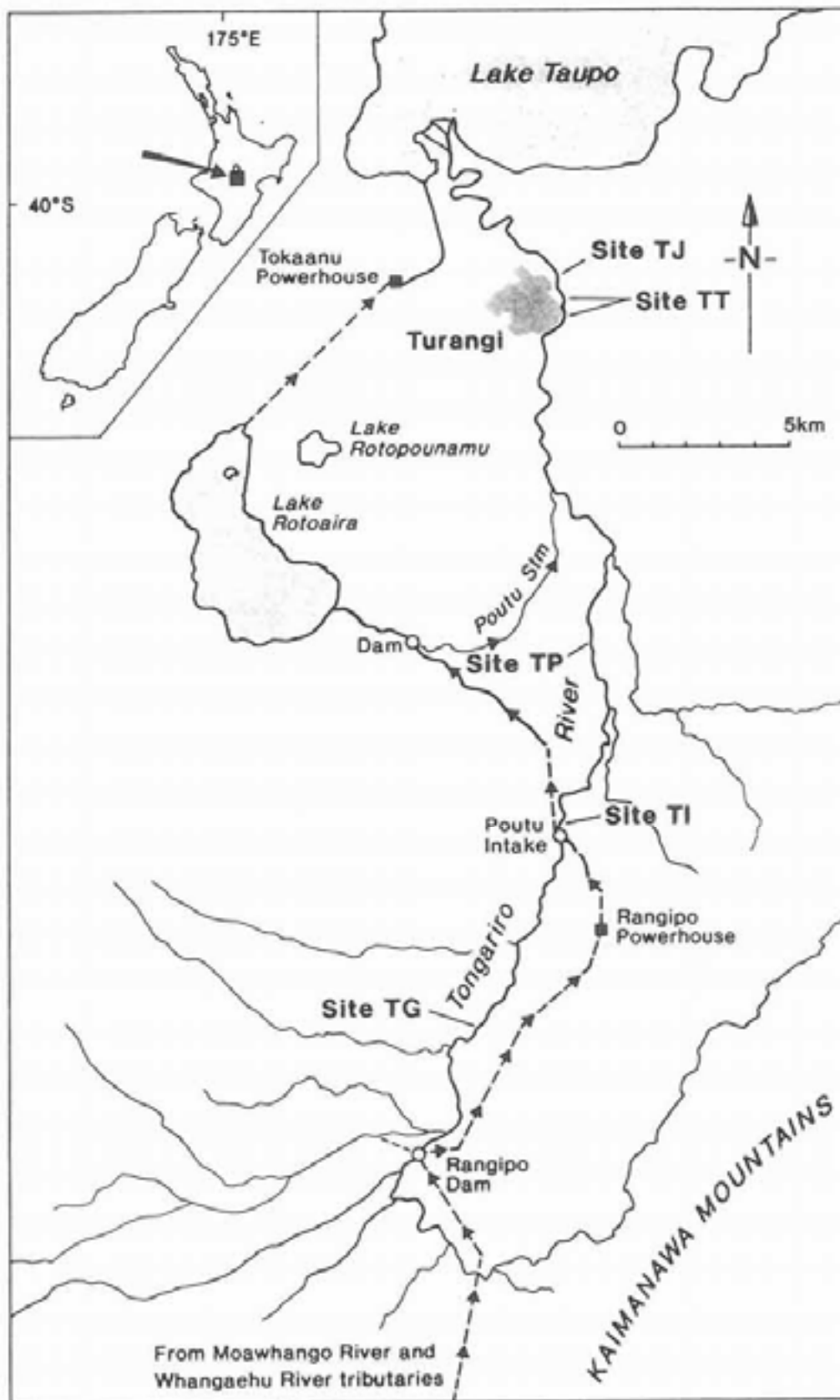


Figure 1. Location of sites on Tongariro River sampled in December 1990 and December 1992. See Table 1 for grid references of TG, TI, TP and TT. Grid reference of TJ is NZMS 260 T19 542 418.



Power Development water consents process, I studied the flow preferences in terms of water depth and velocity of benthic (bottom-dwelling) invertebrates in Tongariro River. Preference curves were developed for the dominant taxa, and variations in preferences with invertebrate size were investigated to enable more accurate preference curves to be defined. It is intended that these findings be linked with hydraulic surveys of the river and with information on the diet of blue duck and juvenile trout to determine the likely effects of different flow regimes on food supplies.

## 2. METHODS

### 2.1 Sampling Sites

Invertebrate samples were collected at four sites on Tongariro River (Fig. 1, see Table 1 for grid references) to obtain an overall picture of invertebrate micro-habitat preferences down the river. The uppermost site (TG) was approximately 500 m above Tree-trunk Gorge (Fig. 1). The section of river incorporating this site (i.e., between Rangipo Dam and Waikato Falls) provides the main habitat for blue duck on Tongariro River. A compensation flow of at least  $0.6 \text{ m}^3 \cdot \text{s}^{-1}$  is maintained below Rangipo Dam. Tributaries entering Tongariro River progressively augment river flow down this section. The largest of these is Oturere Stream which enters the Tongariro approximately 2 km above Tree-trunk Gorge.

Site TI (Fig. 1) was immediately below Poutu Intake and approximately 13 km below Rangipo Dam. A discharge of  $11.3 \text{ m}^3 \cdot \text{s}^{-1}$  is maintained below Poutu Intake and there can be rapid and extreme fluctuations in flow (Stephens 1989). Lake-run rainbow trout are found up the river as far as the intake; low numbers of resident trout occur above the intake. Site TP was about 20 km below Rangipo Dam (Fig. 1). Flow fluctuations are less extreme on this part of the river than below Poutu Intake. The lower most

**Table 1 Grid references, distances below Rangipo Dam, ranges of physical factors assessed and the number of benthic samples collected at 4 smapling sites on the Tongariro River in December 1990**

	<b>TG</b>	<b>TI</b>	<b>TP</b>	<b>TT</b>
Grid reference (NZMS 260 T19)	520 222	541 268	550 330	537 414 537 409
Distance (km)	6	13	20	30
Depth (m)	0.10-1.40	0.10-1.50	0.10-0.68	0.10-1.21
Velocity ( $\text{m} \cdot \text{s}^{-1}$ )	0.08-1.53	0.08-1.53	0.08-1.58	0.11-1.78
Substrate index	4.2-5.9	3.9-5.9	4.1-5.5	4.6-5.8
Embeddedness index	1-4	1-4	1-4	3-4
Periphyton index	2-5	2-4	2-4	2-4
No. of samples	23	26	13	21

sampling site, TT, was adjacent to Turangi township and covered parts of the river between Hydro Pool and Breakfast Pool. A constant baseflow of  $27.7 \text{ m}^3 \cdot \text{s}^{-1}$  is maintained in the Tongariro River at Turangi, but short periods of high flow may occur following storms (Quinn and Vickers 1992).

## 2.2 Sample Collection

Eighty-four Surber samples were collected from the four sites between 11-14 December 1990. December was selected as the sampling month because, at this time, juvenile trout numbers and feeding activity are highest (Stephens 1989), and blue duck pairs usually have broods on the river (Cam Speedy, Department of Conservation, pers. comm.). Thus, early summer represents the time of year that there is likely to be the greatest demand on the invertebrate food resources provided by the river.

Sampling was stratified to encompass cells in a pre-defined matrix of depth and velocity points ranging, respectively, from 0.10 to 1.50 m and 0.08 to  $1.78 \text{ m} \cdot \text{s}^{-1}$  (Table 1). Water depth above each Surber sampling quadrat ( $0.1 \text{ m}^2$  and velocity at  $0.6 \times$  the depth from the water surface were measured using an OSS PC1 meter attached to a graduated rod (Hydrological Services Ltd.).

The percentage cover of different sized substrata in each quadrat was determined through a viewing box. The size classes used were (after Bovee and Milhous 1978): sand (0.06-2 mm), fine gravel (2-32 mm), coarse gravel (32-64 mm), cobbles (64-256 mm) and boulders (>256 mm). Proportional cover was converted into a substrate index as described by Jowett *et al.* (1991). Embeddedness of the substrate and periphyton cover within each Surber quadrat were assessed qualitatively as described in Table 2. Ranges of values measured at each site are given in Table 1. The index for periphyton cover has been shown to be positively correlated with chlorophyll *a* concentration and ash-free dry weight of periphyton in the Tongariro River (Quinn and Vickers 1992).

**Table 2 Descriptors of embeddedness and periphyton indices used in this study**

Score	Embeddedness index	Periphyton index
1	No packing evident, loose assortment easily moved	Clean substrate
2	Mostly a loose assortment with little overlap	Substrate slippery but periphyton growth not visible
3	Moderately packed with some overlap	Growth obvious with green/brown colour
4	Tightly packed and/or overlapping	Filamentous algae obvious, especially on downstream side
5	Bedrock	Filamentous algae covering >80% of upper surface and sides

Invertebrates colonising the substrate to a depth of about 10 cm in each Surber quadrat were brushed into a 0.25 mm mesh net. Where water depth exceeded about 0.8 m, samples were collected using SCUBA and a harness rope attached to upstream anchors to maintain position in fast water. All samples were preserved in 10% formalin.

### **23 Sample Analysis**

In the laboratory, samples were passed through 1 mm and 0.2 mm mesh nested sieves. Material retained by the latter sieve was sorted at 10 X magnification in a Bogorov tray, and invertebrates caught on the 1 mm mesh sieve were picked out by eye. All identification and counting was done at 6-40 X magnification. Aquatic invertebrates were placed into one of eight taxonomic groups for subsequent analyses (see Table 7 for list of major taxa). A complete list of all invertebrate taxa collected in this study is given in Appendix 1.

The microdistributions of some taxa were examined in more detail to provide additional information on flow preferences at different life-history stages and amongst species within the broader taxonomic groupings. These analyses were carried out only at selected sites because of the intensity of labour involved. Larval head widths of the mayfly *Deleatidium* spp. and the net-spinning caddisfly *Aoteapsyche* spp., and pronotum widths of the caddisfly family Hydrobiosidae (excluding the genera *Psilochorema* and *Neurochorema*; see later) collected from TG and TI were measured with an eye-piece graticule. These two uncased caddisfly taxa can be important components of blue duck diet (Veltman *et al.* in prep.). For *Deleatidium* spp., which is relatively more abundant in the upper than in the lower river (Quinn and Vickers 1992), numbers of final instars (i.e., those with fully developed wing-pads) were also noted.

Chironomid (midge) larvae are dominant in the diet of juvenile trout in the Tongariro River during summer (Stephens 1989), and can also comprise a large proportion numerically of blue duck diet (Collier 1991, Wakelin in press). Other work in this river has shown that the chironomid fauna is comprised of up to six dominant species (Quinn and Vickers 1992). In the samples I collected from sites TI, TP and TT, chironomid larvae were sorted into one of 10 taxa (see Appendix 1). Only five of the taxa recognised were abundant and represented 83-99% of the total chironomid fauna (see Table 9 for list of major taxa). Where chironomids were particularly abundant (most samples from sites TP and TT), subsamples (162-882 individuals) were taken for identification using a Folsom splitter. Numbers of different taxa in each subsample were adjusted for the proportion of the total chironomid numbers they represented (range = 13-56 %) to give number per 0.1 m<sup>2</sup> of riverbed. After identification, the biomass of total chironomids collected in each Surber sample at TI, TP and TT was determined by drying larvae overnight at 60°C and weighing them to the nearest 1 mg on an Ohaus Galaxy 400D balance.

### **2.4 Analysis of Site Effects**

Subsets of nine samples collected from the same combination of cells at all sites (depths 0.10-0.42 m, velocities 0.28-1.17 m.s<sup>-1</sup>) were used to investigate gross site-related differences in invertebrate abundance. Samples sizes ranged from 36 (4 sites) for the main invertebrate groups and 27 (3 sites) for the different chironomid taxa.

Multiple pairwise comparisons between sites ( $\log_e$  transformed data) for different invertebrate groups were made with the Bonferroni test in SYSTAT.

## 2.5 Analysis of Preference Curves

Pearson correlation coefficients between measured physical variables (depth, velocity, substrate size index, substrate embeddedness and periphyton cover) were calculated to identify uncorrelated variables that could be used in multiple linear regression analyses with taxonomic richness ( $\log_e$  transformed), density and chironomid biomass ( $\log_e(N+1)$  transformed). All variables were used in regression analyses except for substrate which was correlated with velocity and embeddedness (Table 3). Depth and velocity preference curves were subsequently calculated for those invertebrate groups which were significantly affected by depth or velocity in the regression analyses.

Preference curves were determined using two methods that have been found to be effective in determining flow preferences of aquatic invertebrates in other New Zealand studies (Jowett and Richardson 1990, Jowett *et al.* 1991). In the "Lowess" method, relative abundance was first calculated by dividing the abundance of each taxon in different samples by the maximum abundance of that taxon at each site. This produced "preference factors" ranging from 0 (no larvae collected) to 1 (maximum abundance at a site), and eliminated the tendency for data from one site with high abundances to obscure patterns at other sites with lower abundances (Jowett *et al.* 1991). The same procedure was also carried out on total numbers of invertebrates and the number of taxa, and on chironomid biomass at TI, TP and TT using maximum biomass at each of three sites as the denominator.

Locally weighted smooth ("Lowess") curves were then plotted through the data for each site using the Lowess smoothing routine with a tension of 0.5 in SYSTAT. This routine produces a smooth by running along the x axis and finding predicted values from a weighted average of nearby y values. Optima were considered to occur at the maximum value of the fitted curve. Where there appeared to be more than one maximum value on the curve, the value which corresponded most closely to the maximum of the fitted polynomial curve (see below) was chosen. Generalised Lowess-smoothed curves were derived by combining normalised data from each site.

**Table 3 Pearson correlation coefficients between physical variables measured in Tongariro River (all sites combined).**

	Velocity	Substrate	Embeddedness	Periphyton
Depth	0.01	-0.09	-0.05	-0.11
Velocity		0.37**	0.06	-0.06
Substrate			0.31**	0.15
Embeddedness				0.09

\*\* =  $P < 0.01$ . n = 83

The other method used for calculating preference curves involved deriving second order exponential polynomial regression equations for depth or velocity (Orth and Maughan 1983). For abundance and biomass data, these equations took the form:

$$(1) \log_e(N+1) = a + bX + cX^2$$

where N is density or biomass of invertebrates, X is depth or velocity and a, b and c are fitted coefficients obtained using the MGLH routine of SYSTAT. Log<sub>e</sub> transformations were used to help normalise the data as confirmed by assessment of frequency distributions.

Polynomial preference curves for taxonomic richness were fitted in the same way using log<sub>e</sub> transformed data. Maximum values of density or taxonomic richness were derived from the polynomial regression curves and preference factors ranging from 0 to 1 were determined by dividing the equation by the maximum value estimated by the function in the range of conditions sampled (Orth and Maughan 1983). Generalised exponential polynomial preference curves were derived by combining normalised data from each site.

## 2.6 Analysis of Larval Size Effects

Peaks in size-frequency distributions of taxa measured at sites TG and TI were used to identify classes for determination of the relationship between larval size and microhabitat preferences (Fig. 2). This analysis identified three *Aoteapsyche* spp., four Hydrobiosidae and eight *Deleatidium* spp. size classes (Table 4). Because of the limited size range of *Aoteapsyche* spp. larvae collected from the Tongariro River in December 1990, it was not possible to assess the effects of larval size on microhabitat preferences for this taxon, and these data were not considered further. Preference curves for different size classes of Hydrobiosidae and *Deleatidium* spp. at the two sites were calculated as described above. Larvae of the hydrobiosid genera *Psilochorema* and *Neurochorema*,

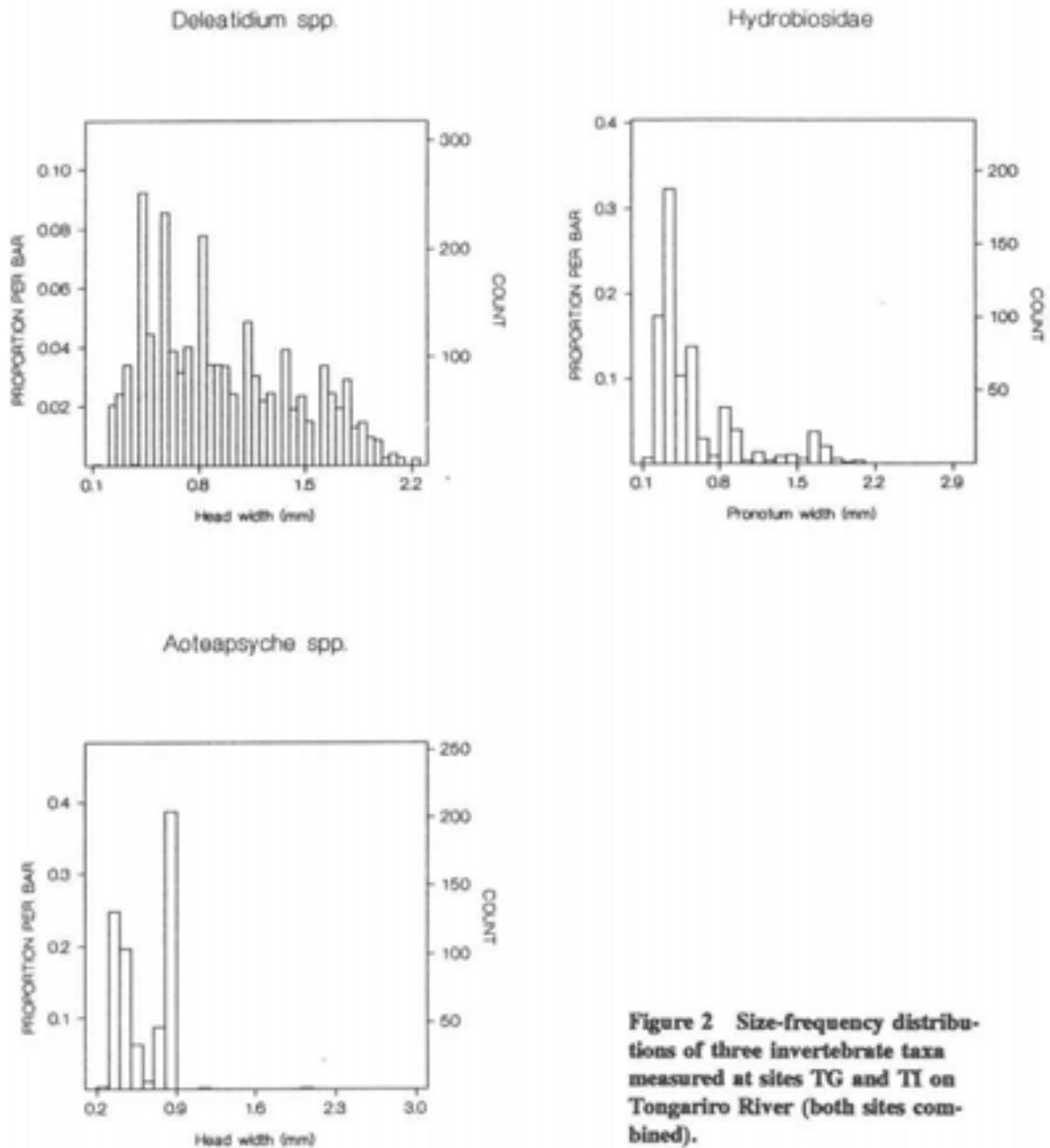
**Table 4 Range of sizes (mm except for final instar (FI) *Deleatidium* spp. which were recognised from fully developed wing pads) for head widths (*Deleatidium* and *Aoteapsyche* spp) and pronotum widths (hydrobiosidae) in different size classes differentiated from size frequency distributions (see Fig. 2). Number of larvae in each size class are given in parentheses.**

Size class	I	II	III	IV	V	VI	VII	VIII
<i>Deleatidium</i> spp.	<0.40 (216)	0.40- 0.50 (373)	0.55- 0.75 (536)	0.80- 1.05 (557)	1.10- 1.30 (341)	1.35- 1.55 (265)	1.6-<F1 (255)	F1 (189)
Hydrobiosidae	<0.45 (317)	0.45- 0.75 (138)	0.8-1.05 (63)	>1.05 (64)	-	-	-	-
<i>Aoteapsyche</i> spp.	<0.65 (270)	0.65- 0.95 (256)	>0.95 (2)	-	-	-	-	-

- not applicable

\* not considered in subsequent analyses

*Deleatidium* spp. Groups I & II, III & IV, V & VI and VII & VIII were combined for log linear modelling (see text for explanation).



**Figure 2** Size-frequency distributions of three invertebrate taxa measured at sites TG and TI on Tongariro River (both sites combined).

which could be distinguished as early instars, were omitted from the Hydrobiosidae group to make it as taxonomically homogenous as practical in order to minimise the effects of interspecific differences in flow preferences. Most larvae in the Hydrobiosidae group appeared to belong to the genera *Hydrobiosis* and *Costachorema*. *Psilochorema* and *Neurochorema* species made up only small proportions (3% and 6%, respectively) of the total Hydrobiosidae.

Because of the low numbers of larvae collected in some size classes, more detailed statistical analysis attaching significance levels to the effects of size was possible only for *Deleatidium* spp. nymphs. Interactions between sampling site, depth and velocity, and size of *Deleatidium* spp. nymphs were investigated using log-linear modelling in

SYSTAT. Log-linear modelling is a form of multi-dimensional contingency table analysis in which the log of frequencies is expressed as a function of main effect (categorical) and interaction parameters. Depth and velocity data were each separated into five groups for this analysis (Table 5). Some fast velocity sampling points did not fall into discrete groups and were omitted for the purposes of log linear modelling. To minimise the number of sparse cells (frequency <5), the size data were compressed into four size classes for analysis by combining classes I and II, III and IV, V and VI and VII and VIII. This reduced the proportion of sparse cells to 25%. This slightly exceeded the recommended 20% level for sparse cells, and so a higher than normal probability level ( $P < 0.01$  instead of  $P < 0.05$ ) was used to determine significance.

**Table 5 Delimiters of depth (m) and velocity (m.s<sup>-1</sup>) groups used for log- linear modelling at 2 sites on Tongariro River**

Depth Groups	Velocity Groups
0.10	0.08-0.13
0.19-0.22	0.28-0.36
0.39-0.48	0.56-0.74
0.58-0.60	0.92-1.18
1.0-1.20	1.32-1.53

A nested hierarchy of main factors (site, depth group, velocity group and size class) and relevant second and third order interactions was established for log-linear modelling, as recommended by Fienberg (1970). Effects of interactions on numbers of nymphs in Surber quadrats were determined by testing the significance of the sum of likelihood ratio chi-square test statistics and degrees of freedom at subsequent stages of the hierarchy. Four cells were sampled at TI but not at TG and these were regarded as structural zeros during log-linear modelling (i.e., they were not included in the analysis).

### 2.7 Validation Sampling

Five replicate Surber samples (0.1 m<sup>2</sup>, 0.25 mm mesh net) were collected from "slow", "medium" and "fast" water habitats (see Table 6) near Judges Pool (Site TJ on Fig. 1), a site not previously sampled in the Tongariro, on 3 December 1992 to validate velocity preferences indicated by the earlier sampling. Water depth and velocity above each Surber quadrat were measured before benthic sampling as described in Section 2.2, using a Scientific Instruments Model 1205 mini meter with a Stewart Stream Gauging counter. Samples were analysed as in Section 2.3, except that material retained by the 0.2 mm mesh sieve was subsampled by half using a Folsom splitter. Differences in densities of different invertebrate groups and size classes of *Deleatidium* spp. and Hydrobiosidae in the three habitat types (slow, medium and fast) were tested using multiple Bonferroni pairwise comparisons following  $\log_e N + 1$  transformation in SYSTAT. Raw data used in these analyses are presented in Appendix 2.

**Table 6** Depth (m) and velocity ( $\text{m}\cdot\text{s}^{-1}$ ) conditions for 5 Surber samples collected from "slow", "medium" and "fast" flowing water at Site TJ (see Fig. 1) on Tongariro River in December 1992.

Sample	"Slow"		"Medium"		"Fast"	
	Depth	Vel.	Depth	Vel.	Depth	Vel.
1	0.28	0.25	0.31	0.57	0.23	1.17
2	0.25	0.28	0.30	0.60	0.22	0.99
3	0.29	0.13	0.24	0.54	0.19	1.11
4	0.18	0.22	0.25	0.63	0.25	1.23
5	0.13	0.25	0.36	0.60	0.28	1.23
$\bar{x}$ ( $\pm$ SE)	0.23 (0.03)	0.23 (0.03)	0.29 (0.02)	0.59 (0.01)	0.23 (0.01)	1.15 (0.10)

### 3. FLOW PREFERENCES OF MAJOR INVERTEBRATE GROUPS

#### 3.1 Site-related Differences

Significantly higher densities of total invertebrates, chironomids (midges) and oligochaetes (worms) were found at sites TP and/or TT than at the upper two sites on the Tongariro River (Fig. 3). Chironomid biomass was also considerably higher at the two lower sites than at Site TI (Fig. 4). Fewer larvae of the crane-fly *Aphrophila neozelandica* were found at Site TG whereas densities of the mayfly *Deleatidium* spp. were significantly lower at TT than at any other site (Fig. 3). Numbers of the stonefly *Zelandobius furcillatus* and the net-spinning caddisfly *Aoteapsyche* spp. were higher at the lower two sites, but these differences were not statistically significant. On average, similar numbers of taxa were found at all sites in the subset of samples used in this analysis.

#### 3.2 Factors Affecting Distribution

The uncorrelated variables depth, velocity, substrate embeddedness and periphyton cover (see Table 3 for correlation coefficients) accounted for 20% of the variation in taxonomic richness at all sites combined, and for between 10% (elmid beetles) and 68% (*Aoteapsyche* spp.) of variation in abundances of the other invertebrate groups (Table 7). Depth or velocity were significant factors in the model for taxonomic richness, chironomid biomass and abundances of all major taxa and total invertebrates except for Elmidae. Embeddedness was also a significant factor affecting density of total invertebrates, *Deleatidium* spp., *Aoteapsyche* spp., *A. neozelandica*, and Oligochaeta, and numbers and biomass of Chironomidae (Table 7). Relative periphyton abundance was significant only for *Deleatidium* spp. and *Aoteapsyche* spp. Depth and velocity preference curves were calculated for all of the invertebrate groups in Table 7 except for Elmidae.



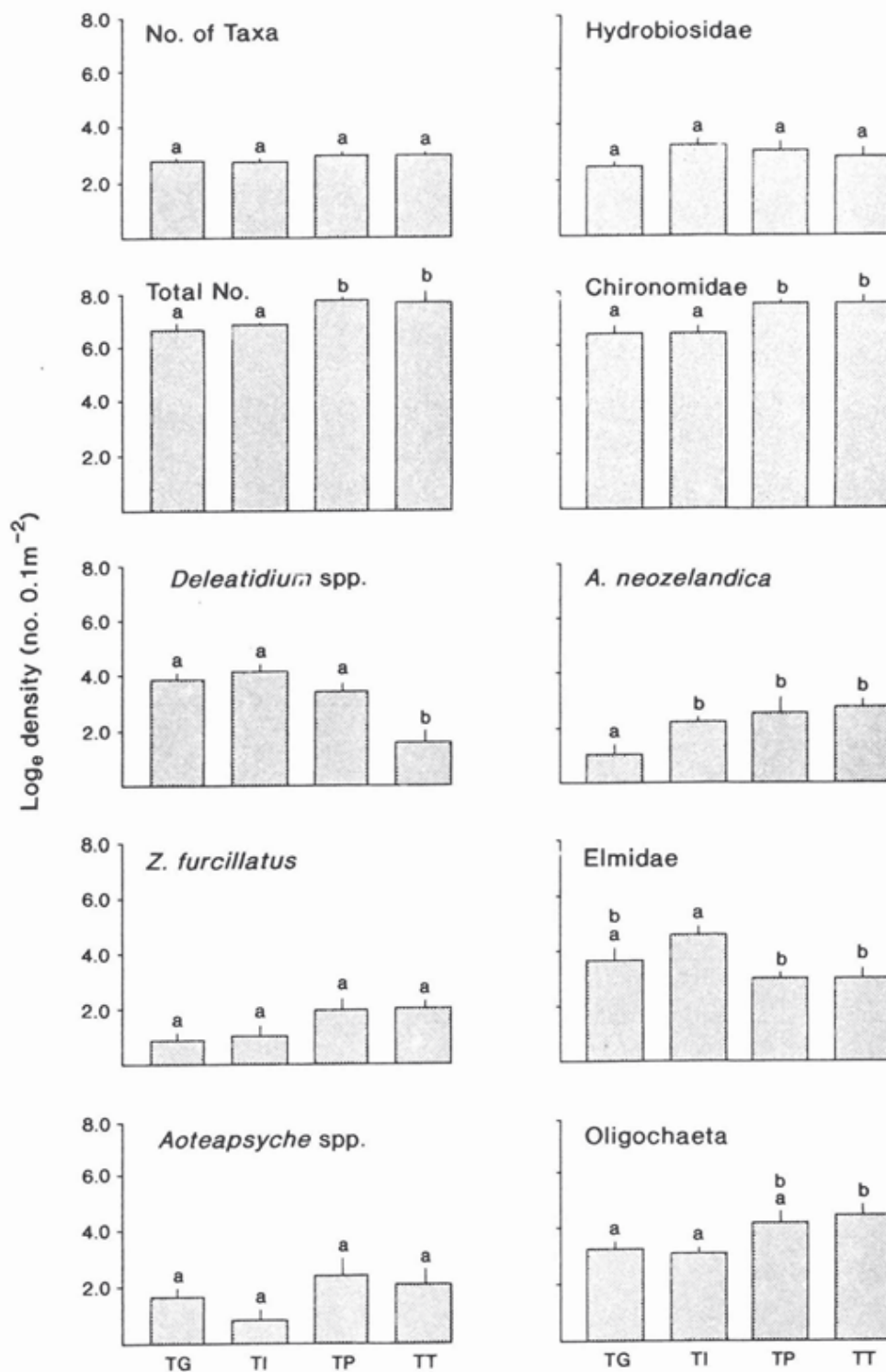


Figure 3 Abundances ( $\log_e N$  or  $N+1$ ,  $x \pm 1\text{SE}$ ) of the major invertebrate groups in subsets of nine samples (see Section 2.4 for explanation) from four sites down Tongariro River in December 1990. Bars with different letters above are significantly ( $P < 0.05$ ) different.

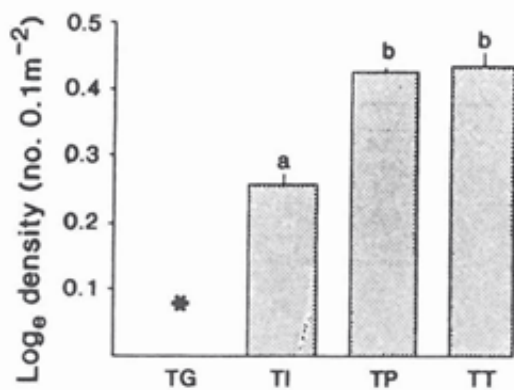


Figure 4 Biomass ( $\log_e N+1, \bar{x} \pm 1SE$ ) of total chironomids in subsets of nine samples (see Section 2.4 for explanation) from three sites down Tongariro River in December 1990. Bars with different letters above are significantly ( $P < 0.05$ ) different. \* = no data.

Coefficients					
Group	Depth	Velocity	Embeddedness	Periphyton	r <sup>2</sup>
Taxa no.	-0.29 **	0.14 *	0.01	0.04	0.20
Total no.	-0.84 ***	-0.14 *	0.31 ***	-0.05	0.34
<i>Deleatidium</i> spp. no.	0.19	1.11 ***	-0.55 ***	0.30 *	0.31
<i>Z. furcillatus</i> no.	-0.56 *	-1.13 ***	0.07	-0.10	0.31
<i>Aoteapsyche</i> spp. no.	-0.86 **	2.71 ***	0.36 **	0.25 *	0.68
Hydrobiosidae spp. no.	-1.24 ***	0.58 *	0.06	0.20	0.29
Chironomidae spp. no.	-0.79 ***	-0.20	0.38 ***	-0.08	0.32
<i>A. neozelandica</i> no.	-0.60 *	0.92 ***	0.46 ***	-0.23	0.35
Elmidae no.	-0.66	-0.26	-0.17	0.18	0.10
Oligochaeta no.	-1.59 ***	-0.93 ***	0.20 *	0.04	0.44
Chironomidae biomass	-1.29 **	0.25	0.43 **	-0.07	0.39

Asterisks indicate probability levels for t statistics associated with the coefficients  
 \* =  $P < 0.05$ ; \*\* =  $P < 0.01$ ; \*\*\* =  $P < 0.001$ .

### 3.3 Flow Preferences of Invertebrate Groups

Preference curves for water depth and velocity are presented in Appendices 3A and B. Similar generalised optima (all sites combined), as indicated by the maximum value of a curve, were obtained for most invertebrate groups using both the Lowess and exponential polynomial curve calculation techniques (see Table 8). Notable exceptions were optimum depths for *Deleatidium* spp. (0.1 m, cf. 1.5 m), and optimum velocities for taxonomic richness, *Deleatidium* spp. and *A. neozelandica* which differed by at least  $0.5\text{m}\cdot\text{s}^{-1}$ . The Lowess technique is more sensitive to outliers than the exponential polynomial method, and this may account for some of the differences observed between the two curve calculation techniques.

Ranges of optima for most invertebrate groups generally differed between sites, although some taxa exhibited reasonably narrow depth and velocity preferences at all

**Table 8** Optimum water depths (m) and velocities ( $\text{m}\cdot\text{s}^{-1}$ ) over the range of conditions sampled for 11 aquatic invertebrate groups at 3 (chironomid biomass) or 4 (all other groups) sites on the Tongariro River. Optima were derived from maxima of locally weighted smooth curves (Lowess) and exponential polynomial curves (Polynomial) (see Appendix 3). Optima are for generalised curves (data from all sites combined: no parentheses) and for curves from each site (range given in parentheses).

	Depth		Velocity	
	Lowess	Polynomial	Lowess	Polynomial
<b>Taxa no.</b>	0.4 (0.1-0.4)	0.1 (0.1-0.4)	1.8 (0.6-1.8)	1.3 (0.1-1.5)
<b>Total no.</b>	0.1 (0.1-0.2)	0.1 (0.1-1.2)	0.3 (0.3-7)	0.1 (0.1-1.5)
<b><i>Deleatidium</i> spp. no</b>	0.1 (0.1-0.9)	1.5 (0.1-1.2)	1.8 (0.6-1.8)	1.1 (0.9-1.4)
<b><i>Z. furcillatus</i> no.</b>	0.1 (0.1-0.6)	0.1 (0.1-1.5)	0.3 (0.1-0.7)	0.1 (0.1-0.6)
<b><i>Aoteapsyche</i> spp. no.</b>	0.6 (0.2-0.6)	0.4 (0.1-0.7)	1.8 (1.3-1.8)	1.8 (1.5-1.8)
<b>Hydrobiosidae no.</b>	0.1 (0.1-0.4)	0.1 (0.1-0.5)	0.3 (0.3-07)	1.0 (0.7-1.8)
<b>Chironomidae no.</b>	0.2 (0.1-0.2)	0.1 (0.1-1.5)	0.3 (0.3-0.7)	0.1 (0.1-1.5)
<b><i>A. neozelandica</i> no.</b>	0.2 (0.1-0.6)	0.4 (0.1-1.3)	0.6 (0.3-1.8)	1.2 (0.4-1.4)
<b>Elmidae no.</b>	0.2 (0.1-0.7)	0.1 (0.1-0.7)	0.1 (0.1-0.7)	0.1 (0.1)
<b>Oligochaeta no.</b>	0.1 (0.1-0.3)	0.1 (0.1-0.3)	0.1 (0.1-0.7)	0.1 (0.1)
<b>Chironomidae biomass</b>	0.2 (0.1-0.2)	0.2 (0.1-0.2)	1.8 (0.6-1.8)	1.8 (0.5-1.8)

Sites (e.g., *Aoteapsyche* spp. and Oligochaeta) (Table 8). Optimum velocities for total chironomid numbers (both curve calculation techniques) and for abundances of *A. neozelandica* and Oligochaeta (Lowess curves only) were higher at TP and TT than at the upper two sites (Appendix 3). In contrast, optimum depths for *Aoteapsyche* spp. and *A. neozelandica* (Lowess curves only) were lower at TP and IT than at the upper two sites. Velocity optima for *Deleatidium* spp., *Z. furcillatus*, *Aoteapsyche* spp., Hydrobiosidae and *A. neozelandica* were always greatest at TT, which had the most stable baseflow compared with other sites (Appendix 3). Depth optima for *Deleatidium* spp. were also highest at TT, but the position of this site amongst depth preference curves for other taxa varied depending on the method used to calculate the curves.

Overall, taxonomic richness, chironomid biomass and densities of most invertebrate groups were greatest at depths of 0.1-0.4 m, although this was variable at different sites for some taxa perhaps reflecting differences in the size of the data sets. Taxonomic richness, *Aoteapsyche* spp., Hydrobiosidae, Oligochaeta and chironomid biomass had limited depth optima at all sites using both techniques (Table 8). The range of depths sampled at site TP was lower than at other sites (maximum depth = 0.68 m), but omission of this site from the calculation of generalised Lowess depth preference curves significantly altered the optimum only for *Z. furcillatus* (from 0.1 m to 0.4 m). Therefore, this site did not introduce any bias for most taxa.

Taxonomic richness and abundances of *Deleatidium* spp. and *Aoteapsyche* spp. were greatest in fast water ( $>1.0 \text{ m.s}^{-1}$ ) when data from all sites were combined, whereas abundances of total invertebrates, *Z. furcillatus*, Chironomidae and Oligochaeta were greatest overall where water velocity was  $<0.5 \text{ m.s}^{-1}$ . In contrast to abundances, generalised velocity optima for total chironomid biomass was  $1.8 \text{ m.s}^{-1}$ , although this differed widely between sites perhaps reflecting differences in velocity optima between taxa (see next section). Generalised optimum velocities for Hydrobiosidae and *A. neozelandica* ranged from 0.6-1.0 and 0.6-1.2  $\text{m.s}^{-1}$ , respectively, depending on the technique used to calculate preference curves. Velocity and depth optima for different size classes of Hydrobiosidae and *Deleatidium* spp. are discussed in Section 5.

The methods used here to estimate optimum depths or velocities provide an indication of where maximum abundance, biomass or taxonomic richness was likely to occur. However, preference curves for some taxa were shallow and did not extend below values of 0.5 (an arbitrarily selected value which indicates approximately where half the maximum density of a taxon occurred). This indicates that these taxa were found over a wide range of conditions and had broad preferences. None of the preference curves calculated using the exponential polynomial method for taxonomic richness and abundances of total invertebrates, chironomids and elmids extended below preference values of 0.5 in the range of depth and velocity conditions sampled for any site (Appendix 3B). The polynomial preference curves for chironomid biomass also did not fall below preference values of 0.5.

### 3.4 Validation Sampling

Replicate sampling of "slow", "medium" and "fast" flowing habitats (see Table 6 for velocities) was carried out at Site TJ (see Fig. 1) in December 1992 to verify

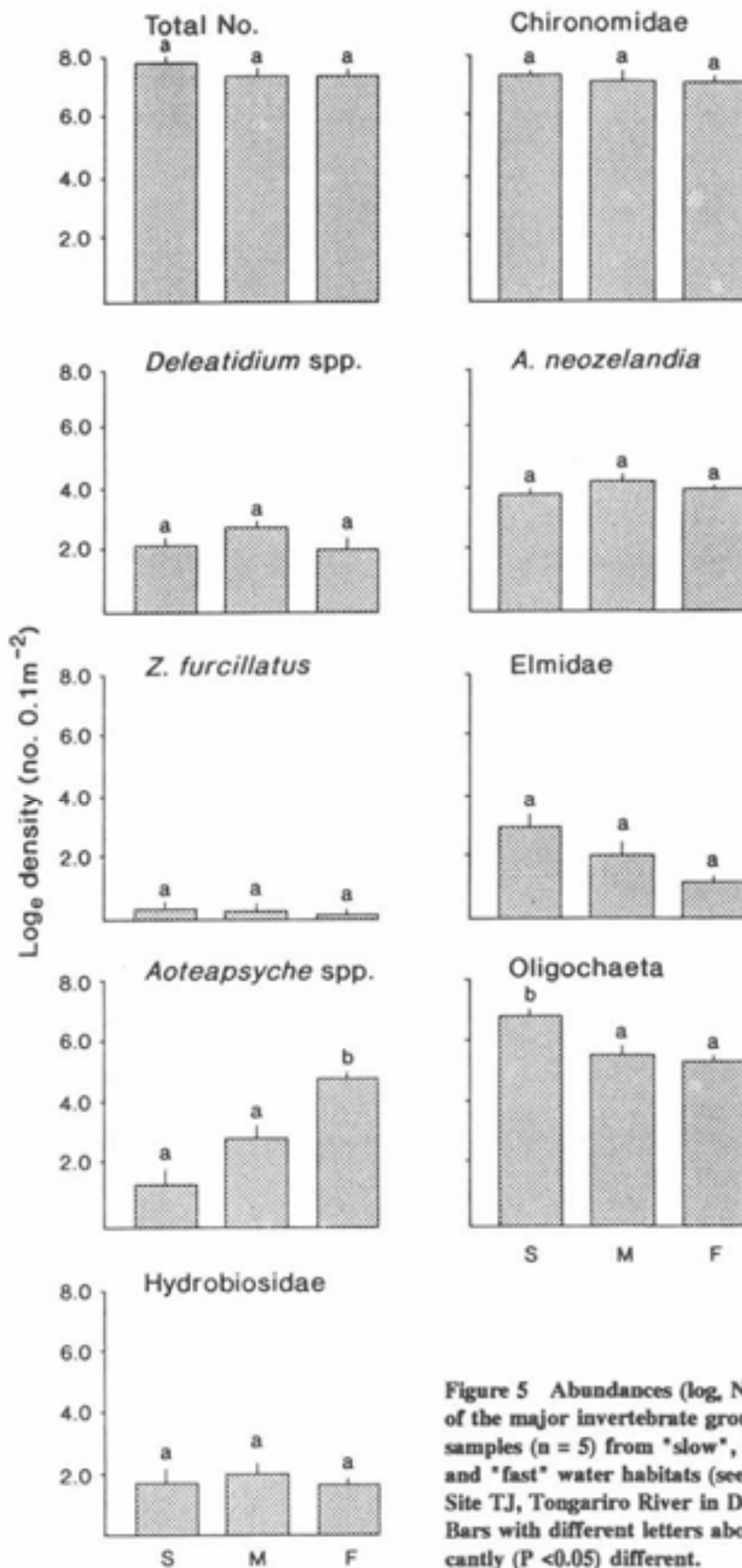


Figure 5 Abundances ( $\log_e N+1$ ,  $\bar{x} \pm 1SE$ ) of the major invertebrate groups in Surber samples ( $n = 5$ ) from "slow", "medium" and "fast" water habitats (see Table 6) at Site TJ, Tongariro River in December 1992. Bars with different letters above are significantly ( $P < 0.05$ ) different.

conclusions derived from preference curve analyses. Densities of total invertebrates, *Deleatidium* spp. and Hydrobiosidae (all size classes combined), *Z. furcillatus*, Chironomidae (all taxa combined), *A. neozelandica* and Elmidae were not significantly ( $P < 0.05$ ) different between the different habitat types (Fig. 5). This result is in general agreement with the preference curve analyses (see Table 8) which indicated broad velocity preferences for most of these invertebrate groups. Densities of the stonefly *Z. furcillatus* were low at TJ in December 1992 and conclusions regarding the slow velocity preference of this taxon could not be verified.

Significant differences in abundances of the uncased caddisfly *Aoteapsyche* spp. and oligochaetes were detected between habitats (Fig. 5). Densities of *Aoteapsyche* larvae increased with mean velocity and were significantly higher in the "fast" flowing habitat (mean =  $1.15 \text{ m.s}^{-1}$ ) than in the "slow" or "medium" flowing habitats. In contrast, densities of oligochaetes were significantly higher in "slow" flowing habitats (Fig. 5). This result confirms that optimum velocities for *Aoteapsyche* and oligochaetes in the Tongariro River are  $>1.0 \text{ m.s}^{-1}$  and  $\leq 0.2 \text{ m.s}^{-1}$ , respectively (see Table 8).

#### 4. FLOW PREFERENCES OF CHIRONOMID TAXA

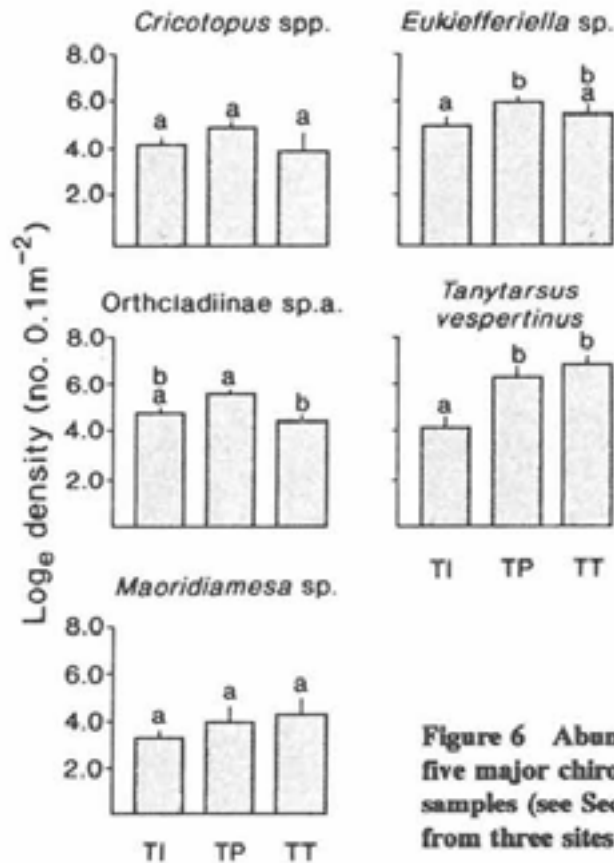
##### 4.1 Site-related Differences

Significantly higher densities of *Eukiefferiella* sp. and *Tanytarsus vespertinus* were found at one or both of the lower two sites on the Tongariro River (Fig. 6). Fewer Orthocladiinae sp.a were found at TT than at TP, but no significant differences in abundances of *Cricotopus* spp. and *Maoridiamesa* sp. were found between any of the three sites examined in this analysis.

**Table 9** Coefficients of multiple regression analyses for the 5 major chironomid taxa using uncorrelated physical variables for all sites combined (see Table 3). Abundance data were  $\log_e(N+1)$  transformed. The percent variance accounted for ( $r^2$ ) by the whole regression is given. Astericks indicate probability levels of t statistic associated with the coefficients.

Coefficients					
Taxon	Depth	Velocity	Embeddedness	Periphyton	$r^2$
<i>Cricotopus</i> spp.	-0.08	0.84 *	0.07	0.36	0.11
<i>Eukiefferiella</i> sp.	-1.16 **	0.75 **	0.20	-0.30	0.39
Orthocladiinae sp. a	-1.75 ***	-0.51 *	-0.14	0.29	0.46
<i>T. vespertinus</i>	-1.23	-1.94 ***	0.94 ***	0.07	0.48
<i>Maoridiamesa</i> sp.	-0.49	2.64 ***	0.34	0.23	0.60

\* =  $P < 0.05$ ; \*\* =  $P < 0.01$ ; \*\*\* =  $P < 0.001$ .



**Figure 6** Abundances ( $\log_e N+1$ ) of the five major chironomid taxa in subsets of 9 samples (see Section 2.4 for explanation) from three sites down Tongariro River.

#### 4.2 Factors Affecting Distribution

The uncorrelated variables depth, velocity, substrate embeddedness and periphyton cover (see Table 3 for correlation coefficients) accounted for between 11% (*Cricotopus* spp.) and 60% (*Maoridiamesa* sp.) of the variation in abundances of five chironomid taxa (Table 9). Water velocity was a significant factor explaining abundances of all major chironomid taxa, whereas depth was important in the multiple regression analysis for *Eukiefferiella* sp. and Orthocladiinae sp.a. Embeddedness was a significant factor affecting abundances of *T. vespertinus* only, and periphyton was not a significant factor for any chironomid taxon (Table 9). Depth and velocity flow preference curves were calculated for all taxa.

#### 4.3 Flow Preferences

Greater resolution of chironomid flow preferences was achieved by calculating individual curves for the five dominant taxa (Appendices 4A and B). Optima of generalised preference curves indicated that most larvae of all taxa occurred predominantly in shallow water ( $\leq 0.6$  m deep; Table 10). When site TP, which was sampled over a narrower depth range than the other sites, was omitted from the analysis of Lowess preference curves, only the generalised depth optimum for *Cricotopus* spp.

was different (from 0.1 to 0.6 m). Thus, inclusion of this site did not introduce any bias for most taxa. Different depth optima were identified for some taxa depending on the site and the technique used to calculate preference curves (Table 10). These differences could partly reflect differences in sample size and in the sensitivity of the techniques used.

When data from all sites were combined, optimum velocities ranged from  $<0.4 \text{ m.s}^{-1}$  for Orthoclaadiinae sp.a and *T. vespertinus*, to  $>1.0 \text{ m.s}^{-1}$  for *Cricotopus* spp., *Eukiefferiella* sp. and *Maoridiamesa* sp. (Table 10). Velocity optima occurred over a wide range at different sites for *Cricotopus* spp. and Orthoclaadiinae sp.a ( $0.1\text{-}1.8 \text{ m.s}^{-1}$ ), but over much narrower ranges for *T. vespertinus* ( $0.1\text{-}0.6 \text{ m.s}^{-1}$ ) and *Maoridiamesa* sp. ( $1.0\text{-}1.8 \text{ m.s}^{-1}$ ). Velocity optima for *Eukiefferiella* sp. at different sites ranged from  $0.6\text{-}1.5 \text{ m.s}^{-1}$  (Table 10).

The methods used here to estimate optimum depths or velocities provide an indication of where maximum abundance is likely to occur. However, preference curves for some taxa were shallow and did not extend below values of 0.5 (an arbitrarily selected value which indicates approximately where half the maximum density of a taxon occurred). This indicates that these taxa were found over a wide range of conditions and had broad preferences. None of the preference curves calculated using the exponential polynomial method fell below preference values of 0.5 for *Eukiefferiella* sp. (depth and velocity), Orthoclaadiinae sp.a (velocity), and *Cricotopus* spp. and *Maoridiamesa* sp. (both depth) in the range of conditions sampled at a site (Appendix 4B). This occurred because these taxa were found over a wide range of conditions at all sites, suggesting that they had broad depth and/or velocity preferences.

**Table 10 Optimum water depths (m) and velocities ( $\text{m.s}^{-1}$ ) for 5 chironomid taxa over the range of conditions sampled at 3 sites on the Tongariro River. Optima were derived from maxima of locally weighted smooth curves (Lowess) and exponential polynomial curves (Polynomial) (see Appendix 4). Optima are for generalised curves (data from all sites combined: no parentheses) and for curves from each site (range given in parentheses).**

Taxon	Depth		Velocity	
	Lowess	Polynomial	Lowess	Polynomial
<i>Cricotopus</i> spp.	0.1 (0.1-0.6)	0.5 (0.1-0.9)	1.8 (0.3-1.8)	1.2 (0.1-1.5)
<i>Eukiefferiella</i> sp.	0.6 (0.1-0.6)	0.4 (0.1-0.6)	1.8 (0.6-1.5)	1.2 (0.7-1.5)
Orthoclaadiinae sp. a	0.2 (0.1-0.6)	0.1 (0.1-0.4)	0.3 (0.3-1.8)	0.1 (0.1-1.5)
<i>T. vespertinus</i>	0.1 (0.1-0.6)	0.1 (0.1-0.2)	0.3 (0.1-0.6)	0.1 (0.1)
<i>Maoridiamesa</i> sp.	0.2 (0.1-0.2)	0.1 (0.1-1.2)	1.8 (1.0-1.8)	1.6 (1.1-1.7)



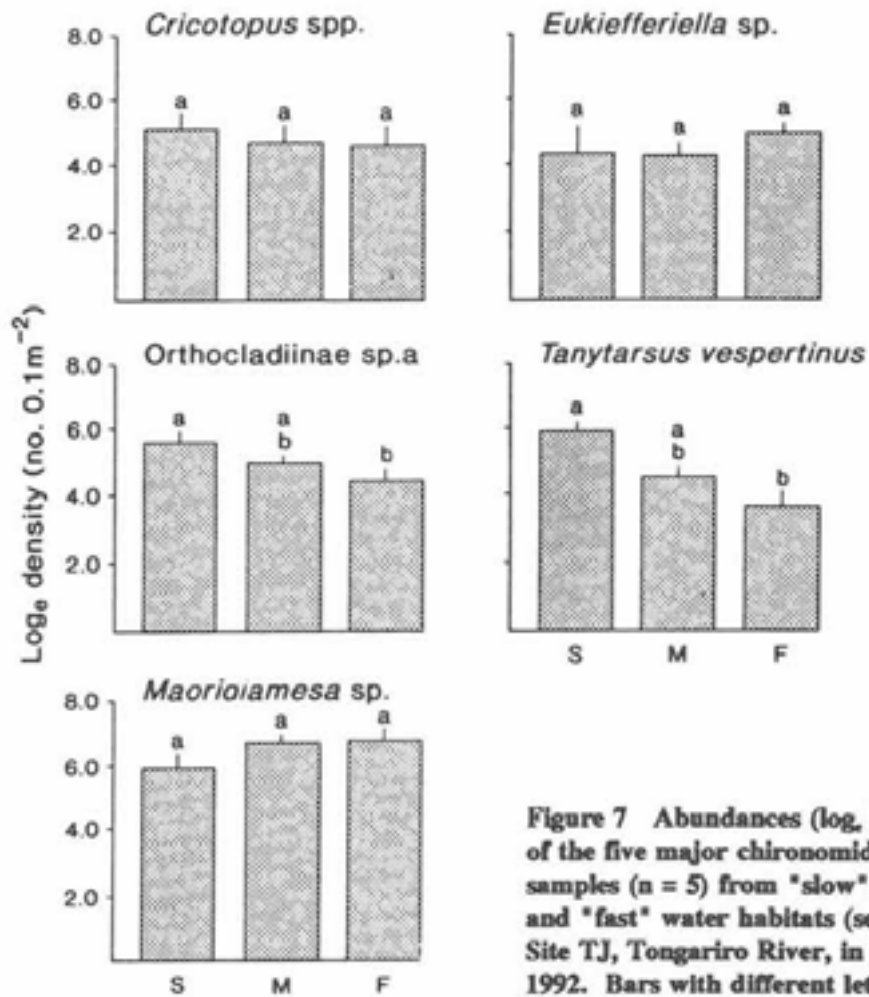


Figure 7 Abundances ( $\log_e N+1$ ,  $\bar{x} \pm 1SE$ ) of the five major chironomid taxa in Surber samples ( $n = 5$ ) from "slow", "medium" and "fast" water habitats (see Table 6) at Site TJ, Tongariro River, in December 1992. Bars with different letters above are significantly ( $P < 0.05$ ) different.

#### 4.4 Validation Sampling

Replicate sampling of "slow", "medium" and "fast" flowing habitats (see Table 6 for velocities) was carried out at TJ (see Fig. 1) in December 1992 to verify conclusions derived from preference curve analyses. Densities of *Eukiefferiella* sp. and *Maoridiamesa* sp. were highest in "fast" flowing habitats, supporting conclusions derived from the preference curve analyses (Fig. 7). However, differences in densities of these taxa between replicate samples at TP were not significantly different, indicating that there was considerable variability between replicates. Some of this variability seemed to stem from the collection of replicate "slow" water samples from the main channel of the river and from a side channel. Densities of *Maoridiamesa* were considerably higher in "slow" flowing habitats in the side channel (703-794 0.1 m<sup>-2</sup>) than in the main river (116-382 m<sup>-2</sup>), indicating that factors other than water velocity can have an effect on densities of this taxon in the Tongariro River.

Densities of *Orthoclaadiinae* sp.a and *T. vespertinus* were significantly higher in "slow" than in "fast" flowing habitats (Fig. 7). This supports conclusions derived from generalised preference curves that these taxa prefer velocities in the range 0.1-0.3 m.s<sup>-1</sup>.

Densities of *Cricotopus* spp. larvae were similar in the different habitats (Fig. 7), confirming the earlier finding that this taxon has broad velocity preferences (see Table 10).

## 5. INFLUENCE OF LARVAL SIZE ON FLOW PREFERENCES

### 5.1 Hydrobiosidae Preferences

Generalised depth optima were similar for each hydrobiosid size class (see Table 4) using both the Lowess and exponential polynomial curve calculation methods (0.1-0.4 m; Table 11). Depth optima for size classes II-IV were different at TG and TI using the exponential polynomial curve calculation technique (Appendix 5B). Thus, optima tended to be deeper (0.6-0.8 m) at TG than at TI (0.1 m).

There was a trend for velocity optima to increase with increasing larval size. Thus, larvae of size classes III and IV had generalised velocity optima of 1.5 m.s<sup>-1</sup> compared with optima of 0.3-1.0 m.s<sup>-1</sup> for size classes I and II, respectively. This trend was consistent for both curve estimation methods even though generalised optima were different for the smallest two size classes (Table 11). At least one of the exponential polynomial preference curves for both sites fell below 0.5 within the range of conditions sampled (Appendix 5B). Log linear analyses were not possible for the hydrobiosid data because of the small number of larvae present in many of the sampling cells.

Replicate sampling of "slow", "medium" and "fast" flowing habitats (see Table 6 for velocities) was carried out at TP (see Fig. 1) in December 1992 to verify conclusions derived from preference curve analyses. Densities of Hydrobiosidae larvae at the time of sampling ranged from 0 to 15 0.lm<sup>-2</sup> for all size classes combined. Densities of

**Table 11 Optimum water depths (m) and velocities (m.s<sup>-1</sup>) over the range of conditions sampled for 4 size classes of Hydrobiosidae at 2 sites on the Tongariro River derived from maxima of locally weighted smooth curves (Lowess) and exponential polynomial curves (Polynomial) (see Appendix 5). Optima are for generalised curves (data from all sites combined; no parentheses) and for curves from each site (range given in parentheses).**

Size Class	Depth		Velocity	
	Lowess	Polynomial	Lowess	Polynomial
Hydrobiosidae I	0.4 (0.2-0.4)	0.1 (0.1)	0.3 (0.3)	0.6 (0.6-0.7)
Hydrobiosidae II	0.1 (0.1-0.4)	0.1 (0.1-0.8)	0.6 (0.6-0.7)	1.0 (0.9-1.5)
Hydrobiosidae III	0.1 (0.1-0.4)	0.1 (0.1-0.7)	1.5 (1.5)	1.5 (1.5)
Hydrobiosidae IV	0.1 (0.1-0.4)	0.1 (0.1-0.6)	1.5 (1.0-1.5)	1.5 (1.2-1.5)

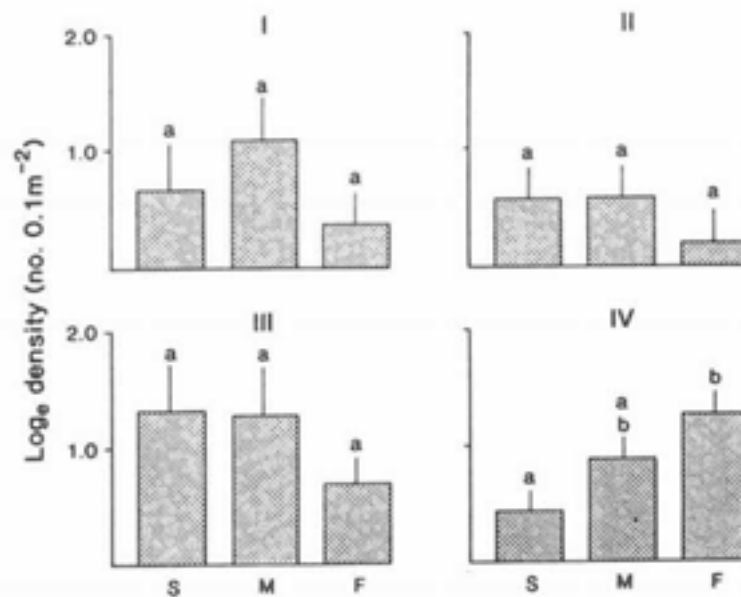


Figure 8 Abundances ( $\log_e N+1$ ,  $\bar{x} \pm 1SE$ ) of the four Hydrobiosidae size classes in Surber samples ( $n = 5$ ) from "slow", "medium" and "fast" water habitats (see Table 6) at Site TJ, Tongariro River in December 1992. Bars with different letters above are significantly ( $P < 0.05$ ) different.

larvae in size classes I-III were not significantly different in "slow", "medium" or "fast" flowing habitats, but a significant difference in the number of size class IV was detected (Fig. 8). Thus, densities of size class IV larvae increased with mean velocity and were significantly higher in "fast" than in "slow" flowing habitats. This supports the general conclusion from the preference curve analysis (Table 11), although the difference in size class III indicated by the preference curve analysis was not evident in the validation sampling.

### 5.2 *Deleatidium* spp. Preferences

Generalised depth preference curves for different size classes of *Deleatidium* spp. indicated that the optimum depth for most nymphs was usually  $< 0.5$  m, although size class VI had a depth optimum of 1.5 m (Table 12, Appendices 6A and 6B). Both curve calculation methods suggested an increase in generalised velocity optima with increasing size up to final instar when velocity optima appeared to decrease. Each site showed this same general pattern, although it was more evident at 11 and the size of *Deleatidium* spp. nymphs at which the velocity optima seemed to increase was different (Appendix 6A and 6B). Generalised curves suggested an increase in velocity optima for nymphs above size class II ( $> 0.5$  mm head width).

The methods used here to estimate optimum depths or velocities provide an indication of where maximum abundance of different size classes is likely to occur. However, preference curves for some taxa were shallow and did not extend below

values of 0.5 (an arbitrarily selected value which indicates approximately where half the maximum density of a taxon occurred). This indicates that these taxa were found over a wide range of conditions and had broad preferences. Velocity preference curves calculated using the exponential polynomial method extended below preference values of 0.5 for all size classes of *Deleatidium* spp. in the range of conditions sampled (Appendix 6B). Because size classes IV-VII were abundant over the range of depths, preferences values did not fall below 0.5 at any site.

Log-linear modelling showed that interactions between site and size class were not significant, indicating that similar numbers of nymphs were present in the same size classes at both sites (Table 13). However, site-related interactions involving size class and depth or velocity were highly significant ( $P < 0.001$ ) suggesting that sampling site had a marked effect on sizes of *Deleatidium* spp. nymphs recorded in different sampling cells. Because significant site effects were detected for both size class analyses, the significance of the effects of depth and/or velocity were tested separately at each site. Effects of depth and/or velocity on numbers of *Deleatidium* spp. in different size classes were highly significant at each site, indicating that these factors affected the size distribution of nymphs (Table 14). Velocity had a stronger effect on

**Table 12 Optimum water depths (m) and velocities ( $\text{m}\cdot\text{s}^{-1}$ ) over the range of conditions sampled for 8 size classes of *Deleatidium* spp. at 2 sites on the Tongariro River derived from maxima of locally weighted smooth curves (Lowess) and exponential polynomial curves (Polynomial) (see Appendix 6). Optima are for generalised curves (data from all sites combined; no parentheses) and for curves from each site (range given in parentheses).**

	Depth		Velocity	
	Lowess	Polynomial	Lowess	Polynomial
I	0.1 (0.1-1.5)	0.1 (0.1-1.5)	0.6 (0.4-0.9)	0.9 (0.8-1.1)
II	0.1 (0.1-1.5)	0.1 (0.1-1.2)	0.6 (0.3-1.0)	0.9 (0.8-1.1)
III	0.1 (0.1-0.4)	0.1 (0.1-1.2)	1.5 (1.1-1.5)	1.2 (0.9-1.5)
IV	0.4 (0.1-0.4)	0.1 (0.1-0.5)	1.5 (1.1-1.5)	1.1 (0.8-1.5)
V	0.6 (0.1-0.6)	0.1 (0.1-0.7)	1.3 (1.1)	1.2 (1.1-1.3)
VI	1.5 (0.4-1.5)	1.5 (0.8-1.5)	1.5 (0.6-1.5)	1.2 (0.9-1.5)
VII	0.4 (0.1-0.4)	0.1 (0.1-1.2)	1.5 (1.5)	1.5 (1.5)
VIII	0.1 (0.1-0.4)	0.4 (0.2-1.5)	1.0 (0.7-1.0)	0.9 (0.8-1.0)

numbers in different size classes at TI than at TG (determined from the chi-square values in Table 14). However, the combined effect of depth and velocity was greater at TG.

Replicate sampling of "slow", "medium" and "fast" flowing habitats (see Table 6 for velocities) was carried out at TJ (see Fig. 1) in December 1992 to verify conclusions derived from preference curve analyses. Although densities of *Deleatidium* spp. nymphs were low at TJ (1-39 O.1m<sup>-2</sup>, all size classes combined), a general trend of greater densities in "fast" flowing water with greater larval size up to final instar was evident (Fig. 9). Differences between habitats were not statistically significant except for size class VII nymphs which reached significantly higher densities in "medium" and "fast" flowing compared with "slow" flowing water. In contrast, final instar nymphs were most abundant in "medium" and "slow" flowing water (Fig. 9) supporting the suggestion that final instars move in to slower flowing habitats, possibly in search of suitable emergence sites. This pattern may not be evident throughout the year as samples were collected in early summer when emergence is likely to be high.

**Table 13 Likelihood ratio chi-square statistics, degrees of freedom (d.f) and probability values (P) for site related effects on numbers of *Deleatidium* spp. in different size classes at two sites on Tongariro River. Size classes I & II, III & IV, V & VI and VII & VIII from Table 4 were combined for log-linear modelling (see Methods).**

Effect	Chi-square	d.f.	P
s*sc	7.4	3	>0.05
s*d*sc	113.4	16	<0.001
s*v*sc	141.3	16	<0.001

**Table 14 Likelihood ratio chi-square statistics, degrees of freedom (d.f) and probability value (P) for depth and velocity related effects on numbers of *Deleatidium* spp. in different size classes at two sites on Tongariro River. Conventions as for Table 13.**

Effect	TG			TI		
	Chi-square	d.f.	P	Chi-square	d.f.	P
d*sc	39.4	12	<0.001	43.8	12	<0.001
v*sc	27.1	12	<0.01	64.4	12	<0.001
d*v*sc	382.3	44	<0.001	300.2	44	<0.001

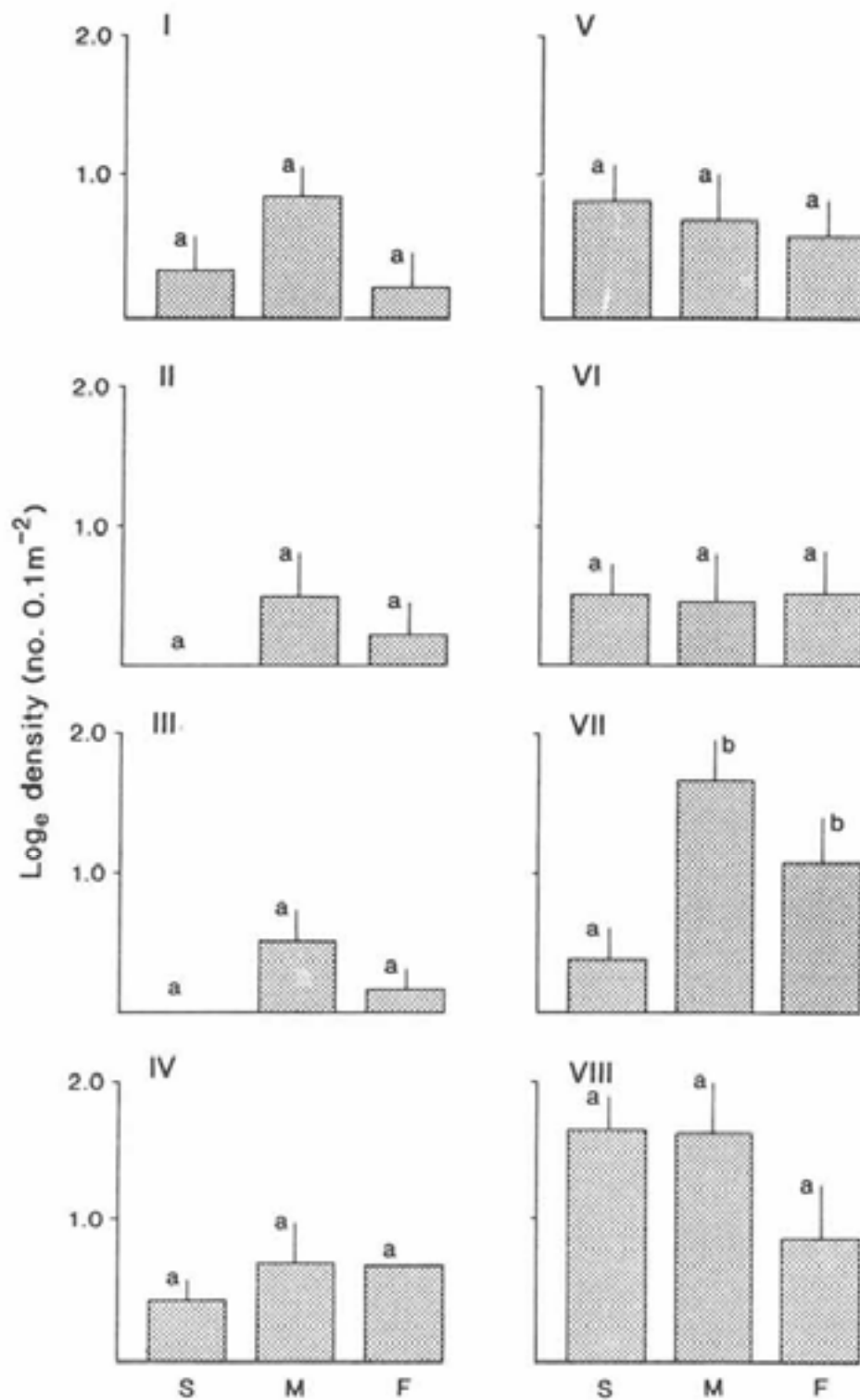


Figure 9 Abundances ( $\log_e N+1$ ,  $\bar{x} \pm 1SE$ ) of the 8 *Deleatidium* spp. size classes in Surber samples ( $n = 5$ ) from "slow", "medium" and "fast" water habitats (see Table 6) at Site TJ, Tongariro River, in December 1992. Bars with different letters above are significantly ( $P < 0.05$ ) different.

## 6. DISCUSSION

### 6.1 Sampling Protocol and Calculation of Flow Preference Curves

The number of samples collected at different sites ranged from 13 (TP) to 26 (TT). It was not possible to sample all cells in the pre-defined matrix of depth and velocity points, and few samples (12 at all sites) were collected in water >0.8 m deep. Thus, the probability of encountering high abundances, biomass or richness of the invertebrate fauna may have been greater in shallow water. However, a good range of velocity samples was taken at all sites (Table 1). The stratified sampling regime minimised the tendency for data to be dominated by a narrow range of depth or velocity conditions that may have prevailed at a specific site. As samples were collected over a narrow time frame (4 days in December), it was not possible to assess whether flow preferences were different in other months.

Both the Lowess and exponential polynomial methods of calculating preference curves yielded similar generalised depth and velocity optima for most invertebrate groups, although there were some differences. Small differences are probably not ecologically significant because of the variable height of different substrate elements influencing depth at a given point in the river, and because there is likely to be a diverse range of flow micro-environments amongst the substrate. However, the two techniques did yield relatively large differences in estimated depth and velocity optima for *Deleatidium* spp. (all sizes combined), depth optima for *Deleatidium* spp. size classes greater than II, and for velocity optima for *A. neozelandica*, *Eukiefferiella* sp. and *Cricotopus* spp. This may reflect the effects of variable sample sizes collected at different sites and the greater sensitivity of the Lowess technique to outliers.

### 6.2 Influence of Site Location

Site location was a significant factor affecting abundance of some of the invertebrate groups examined in the Tongariro River. Densities of all Chironomidae (midge) taxa combined were higher in the lower river sites TP and TT where they comprised 50-92% and 80-97% of total invertebrate numbers, respectively. Biomass of total chironomids, and abundances of *Eukiefferiella* sp. and *T. vespertinus* were also higher in the lower river. Chironomids appear to play an extremely important role in secondary production in the lower river, and this may be particularly so in summer when water temperatures are warmer and development rates are likely to be faster (e.g., Benke and Jacobi 1986). An understanding of the role of chironomids in the energetics of the Tongariro River, and of factors which affect their abundance is therefore important to the development of an ecologically desirable flow regime for the river.

Abundances of total invertebrates, *A. neozelandica*, *Z. furcillatus* and Oligochaeta were also higher in the lower river, but these differences were significant only for the first two-named groups. In contrast, abundances of *Deleatidium* spp. decreased with distance down river. This finding for abundances of total invertebrates, oligochaetes, *Eukiefferiella* sp. and *T. vespertinus* is consistent with the findings of Quinn and Vickers (1992) in summer and in most other seasons at five sites down the Tongariro River (including three sites above TG).

The effect of site location was also evident when deriving site-specific preference curves for some invertebrate groups. For example, the estimated depth optima of total chironomid numbers covered the full range of conditions sampled, perhaps partly reflecting the broad taxonomic composition of this group. In contrast, site location appeared to have a minor effect on depth and velocity preferences of *Aoteapsyche* spp. *Z. furcillatus*, *Oligochaeta*, *Maoridiamesa* sp. and *T. vespertinus* which had similar optima at all sites. Factors leading to apparent site-related effects on flow preferences may include spatial differences in the composition of the taxonomically broader invertebrate groups (see later discussion), the prevailing flow regimes and changes in channel gradient down the river. Velocity optima were highest for several invertebrate taxa (see Appendix 3) at Site TT which had the most stable baseflow, suggesting that prevailing flow regime may have been important in explaining site effects.

### 6.3 Influence of Larval Size

Separation of the mayfly *Deleatidium* spp. and the uncased caddisfly group Hydrobiosidae into size classes provided greater understanding of flow preferences for these taxa which previously have been poorly defined. The finding that different size classes have different velocity optima is supported by the work of Jowett and Richardson (1990) who discovered that dry weights of *Deleatidium* spp. and “uncased caddisflies” (a combination of Hydrobiosidae and *Aoteapsyche* spp.) were positively correlated with water velocity. Water depth and dry weight were correlated positively for uncased caddisflies and negatively for *Deleatidium* spp. in that study, implying that larger caddis larvae occurred predominantly in deeper water and that larger mayfly nymphs were found more commonly in shallower water. Results of the present study suggest that final instar mayflies may move into slower flowing water, possibly in search of suitable emergence sites.

The finding that different size classes of some invertebrate taxa prefer different flow conditions has important implications for the management of invertebrate communities if the availability of suitable habitat is limiting populations. It implies that optimal flow conditions should include a range of water depths and velocities to accommodate different stages of larval growth. It may in fact be the high velocity conditions which are critical because slow water zones are likely to occur in backwaters or behind boulders even at high flows. Because most size classes of *Deleatidium* spp. are usually present in all seasons (Collier and Winterbourn, 1990; Scrimgeour, 1991; Towns, 1981; Winterbourn, 1974), diversity of flow conditions is desirable throughout the year, although sampling in other seasons should be carried out to determine the temporal generality these size-related differences.

### 6.4 Flow Preferences of Invertebrates

In general, taxonomic richness, the density of most invertebrate groups and chironomid biomass were highest in water <0.5 m deep when data for all sites were combined. This is consistent with other work overseas (Orth and Maughan 1983, Brusven 1984) and elsewhere in New Zealand (Jowett and Richardson 1990) which have found that depths around 0.4 m are optimal for most invertebrate groups. Nevertheless, depth did have a significant effect on the distribution of different size classes of *Deleatidium* spp. at TG and TI, and one size class (VI) had an estimated depth optimum of 1.5 m. Few data were collected from this depth though. Depth preference curves for taxonomic richness, and numbers of total invertebrates, total



chironomids, *Cricotopus* spp., *Maoridiamesa* sp., and *Deleatidium* spp. size classes IV-VII were relatively shallow suggesting broad preferences for those groups.

Velocity preferences were much more variable between sites than depth preferences. Taxa could be classified into one of four groups based on their apparent velocity preferences at the different sites determined using both curve calculation techniques (Table 15). Oligochaetes, *Z. furcillatus*, *T. vespertinus* and Hydrobiosidae size class I had slow velocity optima ( $0.1-0.7 \text{ m.s}^{-1}$ ) at all sites, whereas *Eukiefferiella* sp., Hydrobiosidae size class TT and final instar *Deleatidium* spp. consistently preferred "moderate-fast" water velocities (Table 15). "Fast" water taxa (optima  $>0.9 \text{ m.s}^{-1}$ ) included *Aoteapsyche* spp., *Maoridiamesa* sp., the two largest hydrobiosid size classes and most *Deleatidium* spp. nymphs larger than size class II. Validation sampling at site TJ provided support for velocity optima determined for oligochaetes, *Aoteapsyche* spp., *T. vespertinus* and large hydrobiosid larvae.

The two smallest *Deleatidium* spp. size classes were included in the group with "slow-fast" velocity optima ( $0.1-1.8 \text{ m.s}^{-1}$ ), although their estimated optima did not exceed  $1.1 \text{ m.s}^{-1}$ . Other taxa classified as having broad velocity preferences were *A. neozelandica*, *Cricotopus* spp. and Orthoclaadiinae sp.a (Table 15). Jowett *et al.* (1991) made similar conclusions for *A. neozelandica*, and also for oligochaetes and *Aoteapsyche* spp. The shallow exponential polynomial preference curves for taxonomic richness, total invertebrates and total chironomid numbers and biomass suggested that these groups were also relatively common over a wide range of velocity conditions, partly reflecting the broad taxonomic composition of these groups.

### 6.5 Implications for Juvenile Trout Diet

Stephens (1989) found that juvenile trout diet was dominated by chironomid larvae and contained minor proportions of other taxa during summer in Breakaway Pool (approximately 4 km below TP). Furthermore, the stomachs of trout fry were 6 times more full when caught in summer than in winter, indicating that they were feeding more intensively in summer. "Free-living" caddisflies (Hydrobiosidae and *Aoteapsyche* spp.) became important components of trout diet in autumn and winter. The chironomid taxa that were dominant and the size distribution of free-living caddisflies

**Table 15 Groups of aquatic invertebrates divided according to velocity preferences, based on optima at 2-4 sites and calculated using 2 techniques.**

Slow-moderate $0.1-0.7 \text{ m.s}^{-1}$	Moderate-fast $0.6-1.5 \text{ m.s}^{-1}$	Fast $1.0-1.8 \text{ m.s}^{-1}$	Slow-fast $0.1-1.8 \text{ m.s}^{-1}$
<i>Z. furcillatus</i>	<i>Eukiefferiella</i> spp.	<i>Aoteapsyche</i> spp.	<i>Deleatidium</i> I & II
Oligochaeta	Hydrobiosidae II	<i>Maoridiamesa</i> sp.	<i>A. neozelandica</i>
<i>T. vespertinus</i>	Final instar <i>Deleatidium</i> spp.	Hydrobiosidae III & IV	<i>Cricotopus</i> spp.
Hydrobiosidae I		Most III-VII <i>Deleatidium</i>	Orthoclaadiinae sp. a

in the diet of juvenile trout caught by Stephens (1989) are not known. A study of juvenile trout diet at Site TP was carried out in December 1992 and data are currently being analysed by DOC staff at Turangi.

## 6.6 Implications for Blue Duck Diet

Wakelin (in press) examined the contents of seven blue duck faecal samples collected in December 1990 from sites above TG and down to TI on the Tongariro River. He found that, numerically, blue duck diet comprised 19-76% chironomids, 11-49% Trichoptera (a mixture of *Aoteapsyche* spp., Hydrobiosidae and various cased caddisflies), and 2-42% mayflies. *Eukiefferiella* sp. was the numerically dominant chironomid in the diet overall (11%), although *Cricotopus* spp. was relatively abundant in some samples (up to 19% of total invertebrates in the faeces). Chironomids tended to comprise a larger proportion of blue duck diet in samples below than above Tree-trunk Gorge.

The finding that blue duck on the Tongariro River consume a wide range of invertebrate taxa in varying proportions is consistent with the findings of studies on other rivers (Collier 1991, Veltman *et al.*: in prep.). This suggests that the best indicator of food supplies for blue duck may be total abundance and taxonomic richness of the invertebrate fauna. Optima of generalised depth preference curves for total invertebrates and taxonomic richness occurred in shallow water (<0.5 m deep), but velocity optima ranged from 0.1-0.3 and 1.2-1.8 m.s<sup>-1</sup> for total numbers and taxonomic richness, respectively. Nevertheless, preference factors for these groups tended to be high in most depth and velocity conditions sampled (see Appendix 3).

Recent work on Manganuiateao River indicates that blue duck may selectively consume large hydrobiosid larvae (K. Mason, Massey University, pers. comm.), suggesting that it may be necessary to consider larval size when determining flows that should optimise food supplies. If so, suitable habitat would need to be provided for young larvae to ensure that larger ones are recruited to the population. However, Hydrobiosidae represented only a small proportion numerically (7%, all samples combined) of invertebrates in the blue duck faecal samples from the Tongariro River examined by Wakelin (in press).

If total invertebrate abundance and taxonomic richness are the best indicators of blue duck food supplies and they are high over a wide range of flow conditions, as my results suggest, then it may be the area of suitable feeding habitat rather than the amount of food that is limiting blue duck populations on the upper Tongariro. Observations on Manganuiateao River have shown that blue duck feed predominantly in "rippy" or "rippy-white" water (Veltman and Williams 1990) which is probably equivalent to reasonably shallow water with moderately fast current velocities. Observations of blue duck feeding and follow-up measurements of physical conditions are currently being made to define the depth and velocity conditions at which most feeding activity occurs. As with juvenile trout, the influence of blue duck feeding activities on the distribution of invertebrate larvae in the river may also need to be considered when interpreting apparent flow optima for invertebrates.

## 7. CONCLUSIONS

1. In the Tongariro River in December 1990, water depth or velocity were important factors affecting the distribution of all invertebrate groups examined, except for elmid beetle larvae.

2. The different techniques used to estimate depth and velocity optima for invertebrates generally provided similar results. However, there were major differences between techniques for taxonomic richness, *Deleatidium* spp., *A. neozelandica*, *Eukiefferiella* sp. and *Cricotopus* spp. This may reflect the effects of variable sample sizes and the greater sensitivity of the Lowess technique to outliers.

3. Depth and velocity optima varied between sites for some taxa suggesting that other factors were also important in determining distribution. Optima were similar at all sites for the net-spinning caddisfly *Aoteapsyche* spp., the stonefly *Zelandobius furcillatus*, Oligochaeta (worms), and the midge larvae *Tanytarsus vespertinus* and *Maoridiamesa* sp.

4. Depths around 0.4 m are likely to provide suitable habitat for most benthic invertebrate groups and most size classes of Hydrobiosidae and *Deleatidium* spp. Other factors that could influence the decision on appropriate depth conditions in the river should include preferred feeding depths of blue duck, passage requirements of adult trout, and depth requirements for recreation and for stock barriers.

5. Velocity preferences of different taxa varied. Some clearly preferred "slow-moderate" (0.1-0.7 m.s<sup>-1</sup>) velocities (e.g., *Z. furcillatus*, Oligochaeta) whereas others preferred "fast" (1.0-1.8 m.s<sup>-1</sup>) water (e.g., *Aoteapsyche* spp.). Taxonomic richness, chironomid biomass, and abundances of total invertebrates, chironomids, and larvae of the crane fly *Aphrophila neozelandica* were relatively abundant over a broad range of velocity conditions.

6. Separation of chironomid (midge) larvae into major taxa provided considerable insight into the flow preferences of this group which can make up a large proportion numerically of the diet of juvenile trout and blue duck. All chironomid taxa were relatively abundant in shallow water, although *Eukiefferiella* sp., *Cricotopus* spp. and *Maoridiamesa* sp. were relatively abundant over a broad range of depth conditions. Some chironomid taxa showed distinct velocity preferences. *T. vespertinus* occurred predominantly at "slow-moderate" water velocities and *Maoridiamesa* sp. was found mainly in "fast" water. *Eukiefferiella* sp. occurred mostly at "moderate-fast" velocities (0.6-1.5 m.s<sup>-1</sup>) whereas *Cricotopus* spp. appeared to have broad velocity preferences.

7. Velocity preferences of Hydrobiosidae appeared to increase with increasing larval size. Larger *Deleatidium* spp. nymphs also seemed to prefer faster water, although final instar larvae appeared to move into slower water. This suggests that a range of

velocity conditions need to be provided to accommodate different life stages of some invertebrate taxa, at least in early summer.

8. A wide range of invertebrate taxa are eaten by blue duck in varying proportions suggesting that total invertebrate abundance and taxonomic richness may be the best indicators of blue duck food supplies. Total abundance and taxonomic richness were high over a wide range of flow conditions in this study. If these are appropriate indices of blue duck food supplies, then it may be the area of suitable feeding habitat rather than the amount of food that is limiting blue duck populations on the upper Tongariro. A study is currently investigating the water depths and velocities preferred by blue duck for feeding.

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## APPENDIX 1

Taxonomic list of aquatic invertebrates found at the 4 sampling sites on  
Tongariro River in December 1990. Chironomids were separated into taxa at  
sites TI, TP and TT.

+ = present, - = not recorded, \* = not separated.

	TG	TI	TP	TT
<b>Ephemeroptera (mayflies)</b>				
<i>Deleatidium</i> spp.	+	+	+	+
<i>Austroclima jollayae</i>	+	+	+	-
<i>A. sepia</i>	-	+	+	+
<i>Acanthophlebia cruentata</i>	-	+	-	-
<i>Maiulus luma</i>	-	+	+	-
<i>Zephlebia ?spectabilis</i>	-	+	-	-
<i>Zephlebia</i> sp.	-	+	-	-
<i>Nesameletus</i> sp.	+	+	-	-
<i>Coloburiscus humeralis</i>	+	+	+	+
<b>Plecoptera (stoneflies)</b>				
<i>Stenoperla ?maclellani</i>	+	+	-	-
<i>Austroperla cyrene</i>	+	+	+	-
<i>Megaleptoperla grandis</i>	+	+	+	+
<i>Zelandoperla decorata</i>	+	+	+	+
<i>Zelandoperla</i> sp.	+	+	+	-
<i>Zelandobius furcillatus</i>	+	+	+	+
<i>Acroperla trivacuata</i>	+	+	+	+
<i>A. spiniger</i>	-	-	+	-
<i>Halticoperla viridans</i>	+	+	-	-
<i>Spaniocerca zelandica</i>	+	-	-	-
<i>Spaniocerca</i> sp.	-	-	+	-
<b>Trichoptera (caddisflies)</b>				
<i>Aoteapsyche colonica</i>	+	-	-	+
<i>A. ?tepoka</i>	+	+	+	+
<i>A. raruraru</i>	+	+	+	-
<i>Aoteapsyche</i> spp.	+	+	+	+
<i>Orthopsyche fimbriata</i>	-	-	-	+
<i>Oxyethira albiceps</i>	+	+	+	+
<i>Paroxyethira hendersoni</i>	+	-	-	-
<i>Psilochorema</i> sp.	+	+	+	+
<i>Costachorema ?callista</i>	+	+	+	+
<i>C. ?brachyptera</i>	+	+	+	-
<i>C. ?xanthoptera</i>	+	+	+	+
<i>C. ?psaroptera</i>	-	-	+	-
<i>Hydrobiosis parumbripennis</i>	+	+	+	+
<i>H. ?frater</i>	+	+	+	-
<i>H. ?harpidosa</i>	+	+	+	-
<i>H. ?spatulata</i>	+	-	+	-
<i>H. ?charadraea</i>	-	-	-	+
<i>H. ?clavigera</i>	-	-	-	+

[Continued next page]>>

	TG	TI	TP	TT
<b>Trichoptera (caddisflies) (Continued)</b>				
<i>Hydrobiosis</i> sp.A	+	-	-	-
<i>Hydrobiosis</i> sp.B	-	-	+	-
<i>Neurochorema confusum</i>	+	+	+	+
<i>Neurochorema</i> sp.	+	+	+	+
<i>Tiphobiosis</i> sp.	-	+	-	-
Hydrobiosidae	+	+	+	+
<i>Plectrocnemia maclachlani</i>	+	+	-	-
<i>Pseudoconesus</i> sp.	-	+	-	-
Ooconesidae sp.	-	+	-	-
<i>Hudsonema aliena</i>	-	+	-	-
<i>Triplectides ?dolichos</i>	+	-	-	-
<i>Zelolessica cheira</i>	-	+	-	+
<i>Pycnocentrella eurensis</i>	-	+	-	-
<i>Pycnocentroides</i> spp.	+	+	+	+
<i>Beraeoptera roria</i>	+	+	+	+
<i>Confluens hamiltoni</i>	-	+	-	-
<i>Olinga feredayi</i>	+	+	+	+
<i>Pycnocentria funerea</i>	+	+	-	+
<i>P. evecta</i>	+	+	-	-
<b>Coleoptera (beetles)</b>				
Elmidae	+	+	+	+
Hydraenidae	-	+	+	+
Hydrophilidae	-	-	-	+
Helodidae	-	+	-	-
? <i>Liodessus plicatus</i>	-	-	+	-
<b>Diptera (two-winged flies)</b>				
<i>Cricotopus</i> spp.	*	+	+	+
<i>Eukiefferiella</i> sp.	*	+	+	+
Orthoclaadiinae sp.a	*	+	+	+
Orthoclaadiinae sp.A	*	+	+	+
Orthoclaadiinae sp.B	*	+	+	+
<i>Tanytarsus vespertinus</i>	*	+	+	+
<i>Polypedilum</i> sp.	*	-	+	+
Macropelopiini sp.	*	+	+	+
<i>Maoridiamesa</i> sp.	*	+	+	+
Diamesinae sp.	*	+	+	+
Chironomidae indet.	+	+	+	+
Ceratopogonidae	-	+	+	+
Stratiomyidae	-	-	+	-
Empididae sp.A	+	+	+	+
Empididae sp.B	-	-	-	+
Muscidae sp.A	+	+	+	+
Muscidae sp.B	-	+	-	-
Psychodidae	+	-	-	-
Tanyderidae	-	-	+	-
<i>Limonia</i> sp.	-	-	+	-

[Continued next page]>>



	TG	TI	TP	TT
<b>Diptera (two-winged flies) (Continued)</b>				
<i>Aphrophila neozelandica</i>	+	+	+	+
<i>Paralimnophora skusei</i>	+	+	+	+
Eriopterini sp.A	+	+	+	+
Eriopterini sp.B	+	+	+	+
<i>Molophilus</i> sp.	-	+	-	-
Culicidae	-	-	+	-
<i>Austrosimulium</i> sp.	-	+	+	+
<i>Ephydrella</i> sp.	-	-	+	-
<b>Megaloptera (dobsonflies)</b>				
<i>Archichaulioides diversus</i>	-	+	-	+
Oligochaeta (worms)	+	+	+	+
Nematoda (nematodes)	+	+	+	+
Gordiidae (horse hair worms)	+	-	-	-
<i>Potamopyrgus antipodarum</i> (snails)	-	-	-	+
<b>Total no. of taxa<sup>1</sup></b>	<b>50</b>	<b>61</b>	<b>52</b>	<b>43</b>
<b>No. of samples</b>	<b>23</b>	<b>26</b>	<b>13</b>	<b>21</b>

<sup>1</sup> does not include separate chironomid taxa.

## APPENDIX 2

Densities (no.  $0.1\text{m}^{-2}$ ) of invertebrate larvae collected in replicate Surber samples from "slow" (mean velocity =  $0.23\text{ m.s}^{-1}$ ), "medium" (mean velocity =  $0.59\text{ m.s}^{-1}$ ) and "fast" (mean velocity =  $1.15\text{ m.s}^{-1}$ ) flowing water at TJ (see Fig. 1) in December 1992.

"Slow" flowing water.	Sample #	1	2	3	4	5
<b>Ephemeroptera (mayflies)</b>						
<i>Deleatidium</i> spp.		4	11	17	3	11
<i>Zephlebia</i> sp.		-	1	2	-	2
<i>Coloburiscus humeralis</i>		-	-	2	-	11
<b>Plecoptera (stoneflies)</b>						
<i>Zelandobius furcillatus</i>		-	-	2	-	1
<b>Trichoptera (caddisflies)</b>						
<i>Aoteapsyche</i> spp.		-	4	6	1	19
<i>Oxyethira albiceps</i>		3	5	-	40	36
<i>Paroxyethira hendersoni</i>		-	1	-	-	-
<i>Psilochorema</i> sp.		-	-	-	-	4
"Other" Hydrobiosidae		9	10	6	-	8
<i>Helicopsyche</i> sp.		-	1	-	-	-
<i>Pycnocentroides</i> spp.		3	11	15	2	3
<i>Olinga feredayi</i>		-	-	1	-	-
<i>P. evecta</i>		-	1	-	-	1
<b>Coleoptera (beetles)</b>						
Elmidae		52	30	29	4	10
<b>Diptera (two-winged flies)</b>						
<i>Cricotopus</i> spp.		100	248	108	352	228
<i>Eukiefferiella</i> sp.		60	86	70	70	78
Orthocladiinae sp.a		264	206	212	436	472
<i>Tanytarsus vespertinus</i>		1016	562	454	126	174
<i>Maoridiamesa</i> sp.		116	324	382	794	703
Chironomidae indet.		60	18	26	32	36
Empididae sp.		-	3	-	12	20
Muscidae sp.		-	2	2	-	1
<i>Aphrophila neozelandica</i>		25	44	47	24	77
Eriopterini sp.		-	-	1	-	-
<i>Ephydrella</i> sp.		-	-	1	-	-
<b>Megaloptera (dobsonflies)</b>						
<i>Archichaulioides diversus</i>		-	-	-	-	8
Oligochaeta (worms)		1276	1539	373	1203	481

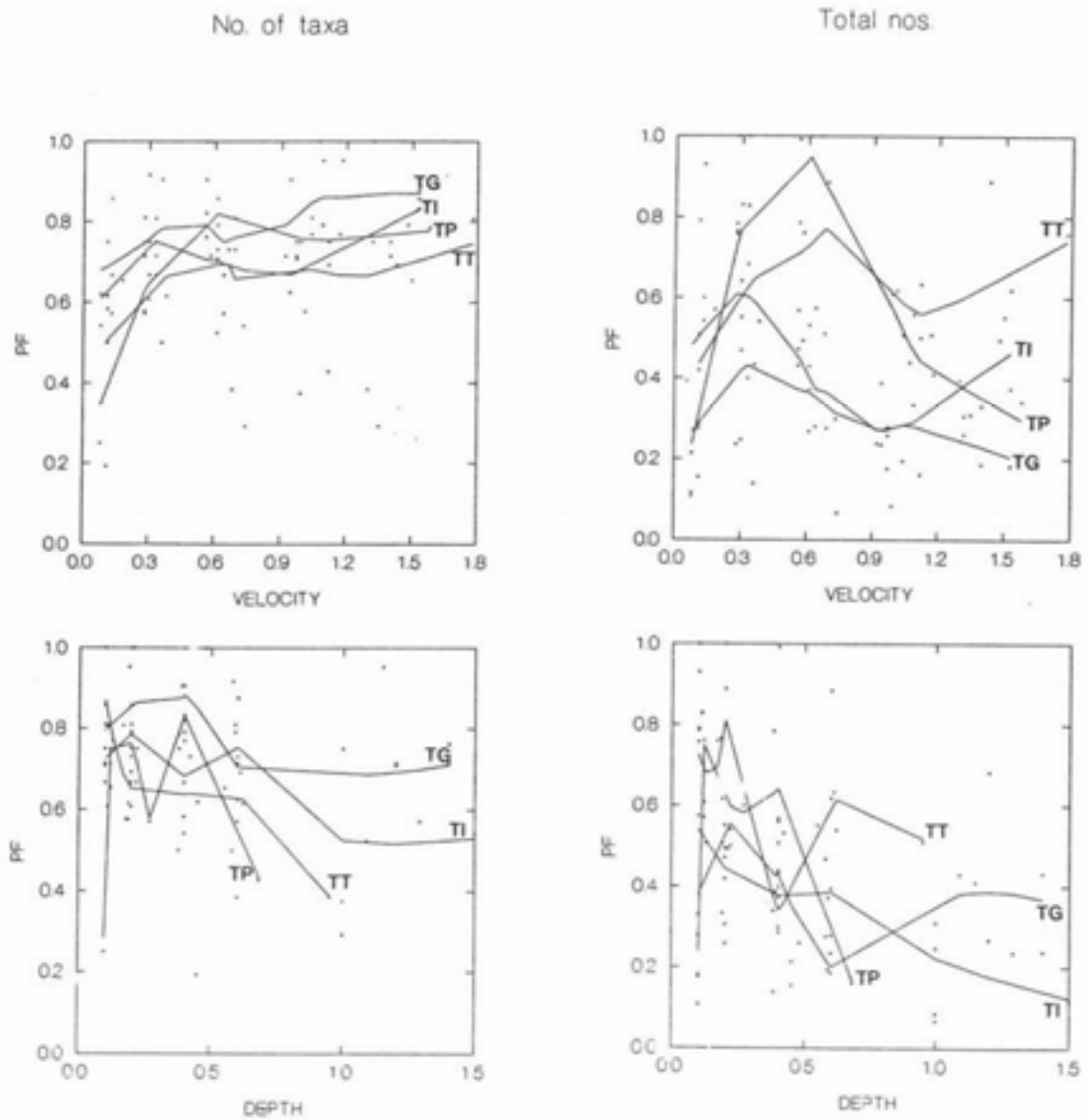
[Continued next page]>>

"Medium" flowing water.	Sample #	1	2	3	4	5
<b>Ephemeroptera (mayflies)</b>						
<i>Deleatidium</i> spp.		12	39	16	9	12
<i>Coloburiscus humeralis</i>		7	1	13	–	3
<b>Plecoptera (stoneflies)</b>						
<i>Zelandobius furcillatus</i>		–	1	1	–	–
<i>Halticoperla viridans</i>		1	–	–	–	–
<b>Trichoptera (caddisflies)</b>						
<i>Aoteapsyche</i> spp.		13	6	60	9	34
<i>Oxyethira albiceps</i>		4	–	4	–	–
"Other" Hydrobiosidae		15	12	6	1	7
<i>Hudsonema aliena</i>		–	–	–	–	1
<i>Helicopsyche</i> sp.		1	–	–	–	–
<i>Pycnocentrodes</i> spp.		10	74	2	–	–
<i>Pycnocentria funerea</i>		–	–	–	–	1
<b>Coleoptera (beetles)</b>						
Elmidae		25	30	5	–	2
Hydrophilidae		1	–	–	–	–
Ptylodactylidae sp.		1	–	–	–	–
<b>Diptera (two-winged flies)</b>						
<i>Cricotopus</i> spp.		102	34	300	110	123
<i>Eukiefferiella</i> sp.		78	22	104	66	104
Orthocladiinae sp.a		104	60	302	126	222
<i>Tanytarsus vespertinus</i>		60	38	212	110	132
<i>Maoridiamesa</i> sp.		990	326	994	758	860
Chironomidae indet.		16	10	38	16	18
Ceratopogonidae		–	–	–	1	–
Empididae sp.		4	–	2	–	2
Muscidae sp.		1	3	6	2	1
<i>Aphrophila neozelandica</i>		84	36	100	50	52
<i>Austrosimulium</i> sp.		–	–	1	–	–
<i>Ephydrella</i> sp.		–	1	–	–	–
<b>Megaloptera (dobsonflies)</b>						
<i>Archichauliodes diversus</i>		–	–	5	1	–
Oligochaeta (worms)		167	217	732	122	268

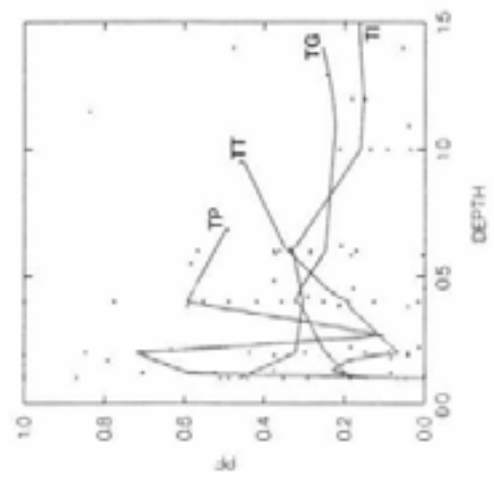
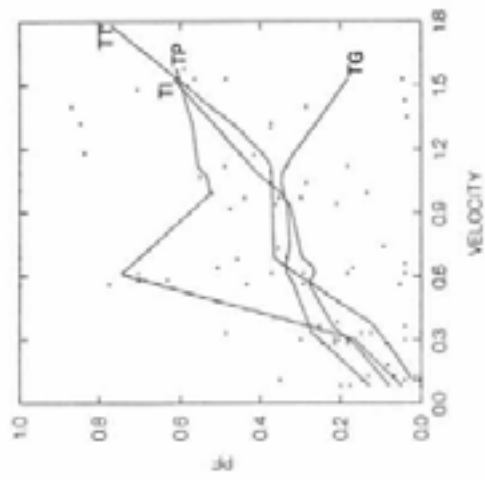
"Fast" flowing water.	Sample #	1	2	3	4	5
<b>Ephemeroptera (mayflies)</b>						
<i>Deleatidium</i> spp.		11	19	11	2	1
<i>Austroclima</i> sp.		-	-	-	1	-
<i>Zephlebia</i> sp.		1	-	-	-	-
<i>Coloburiscus humeralis</i>		10	10	13	9	6
<b>Plecoptera (stoneflies)</b>						
<i>Zelandobius furcillatus</i>		-	1	-	-	-
<i>Acroperla trivacuata</i>		1	-	-	-	-
<i>Spaniocerca</i> sp.		2	-	-	-	-
<b>Trichoptera (caddisflies)</b>						
<i>Aoteapsyche</i> spp.		86	63	132	179	175
<i>Oxyethira albiceps</i>		-	-	4	-	1
"Other" Hydrobiosidae		6	8	4	4	1
<i>Bereoptera roria</i>		-	1	-	-	-
<i>P. evecta</i>		1	-	-	-	-
<b>Coleoptera (beetles)</b>						
Elmidae		2	5	3	2	-
<b>Diptera (two-winged flies)</b>						
<i>Cricotopus</i> spp.		120	126	122	84	53
<i>Eukiefferiella</i> sp.		184	120	180	108	67
Orthoclaadiinae sp.a		116	84	118	142	29
<i>Tanytarsus vespertinus</i>		42	34	78	56	9
<i>Maoridiamesa</i> sp.		860	830	1108	778	430
Chironomidae indet.		44	12	2	20	20
Empididae sp.		-	-	2	1	1
Muscidae sp.		5	1	1	-	1
<i>Aphrophila neozelandica</i>		42	49	69	47	42
<i>Austrosimulium</i> sp.		1	-	-	-	-
<b>Megaloptera (dobsonflies)</b>						
<i>Archichaulioides diversus</i>		-	3	-	-	1
Oligochaeta (worms)		443	159	324	157	117

### APPENDIX 3A

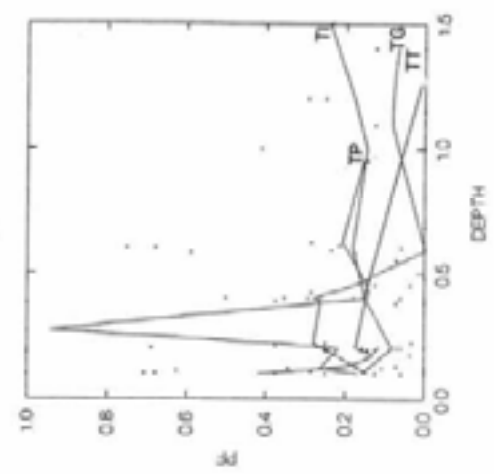
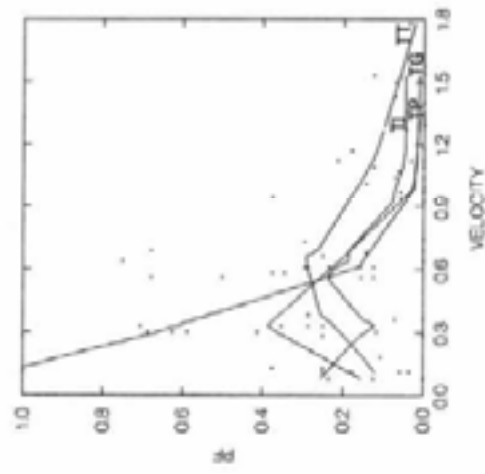
Locally weighted smooth curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for taxonomic richness (number of taxa), total invertebrate densities, densities of 7 major taxa and chironomid biomass in relation to water velocity ( $\text{m.s}^{-1}$ ) and depth (m) at 3–4 sites on Tongariro River.



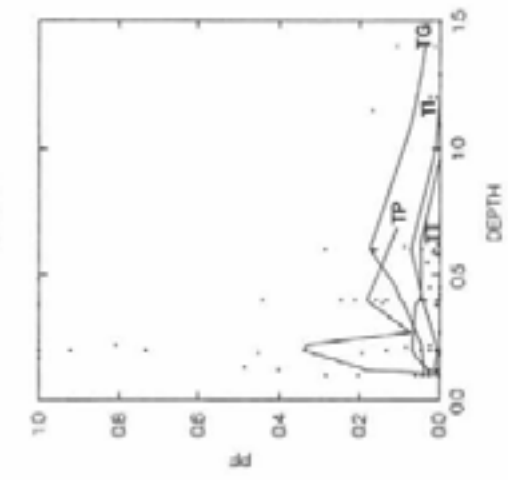
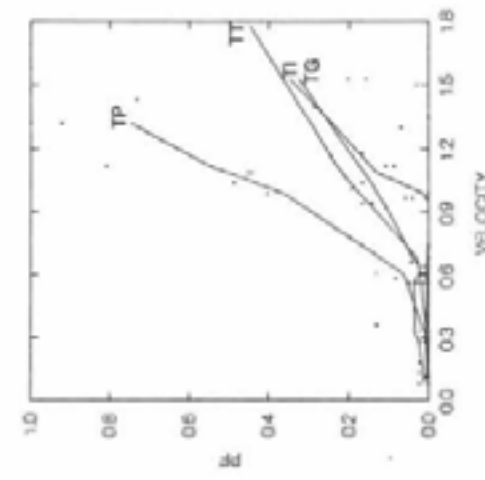
*Deleatidium* spp.



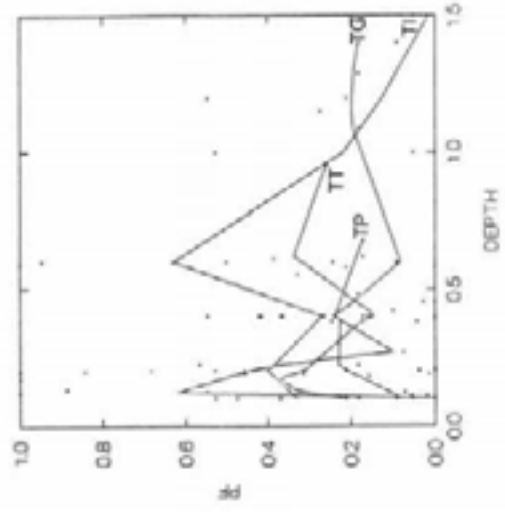
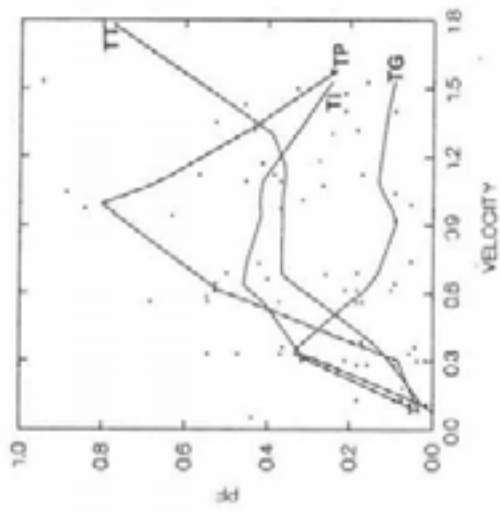
*Z. furcillatus*



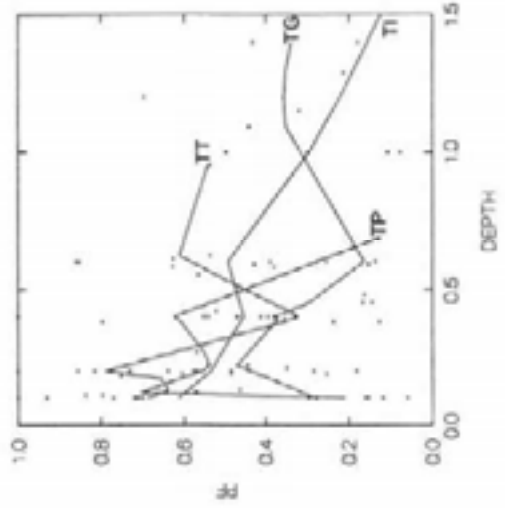
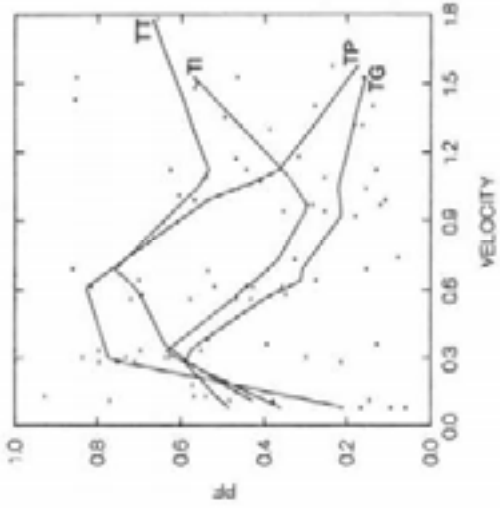
*Aoteapsyche* spp.



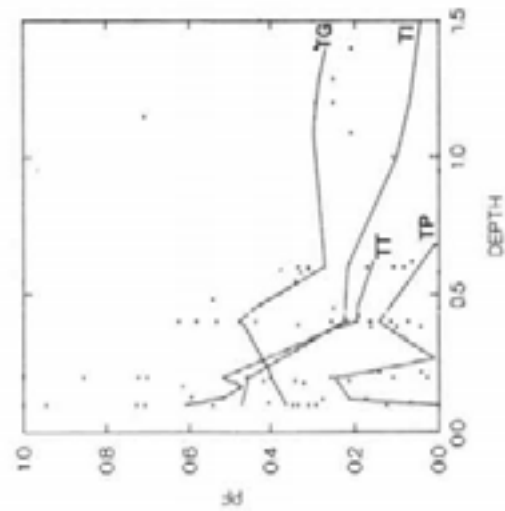
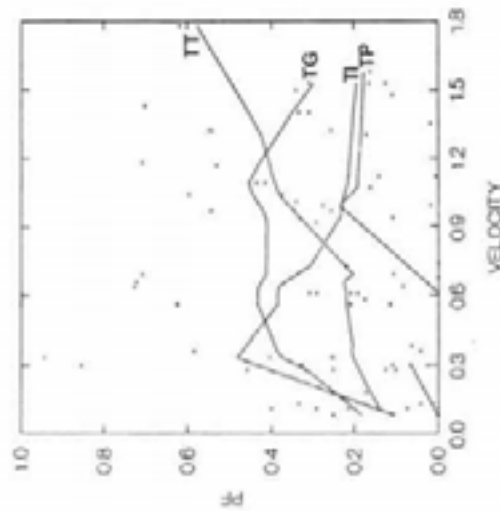
A. neozelandica



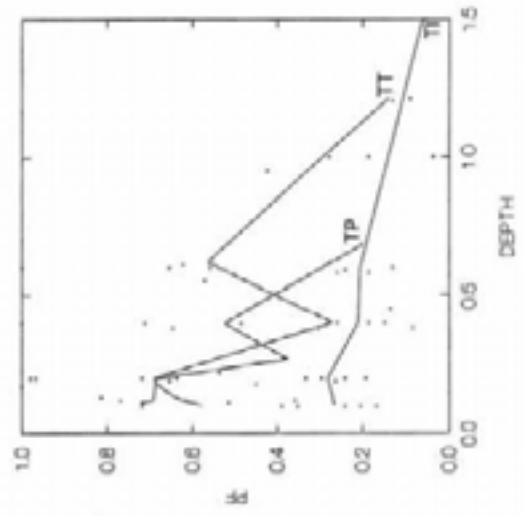
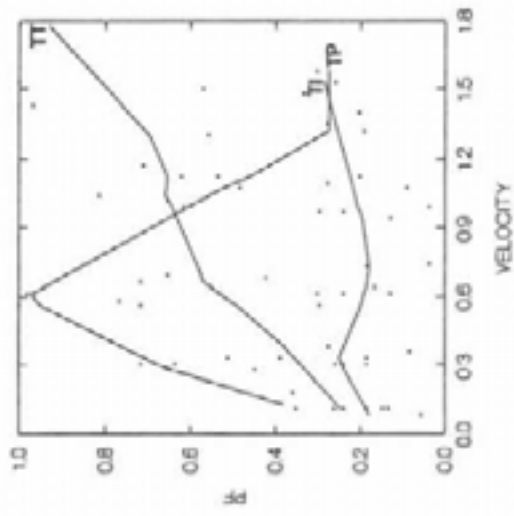
Chironomidae



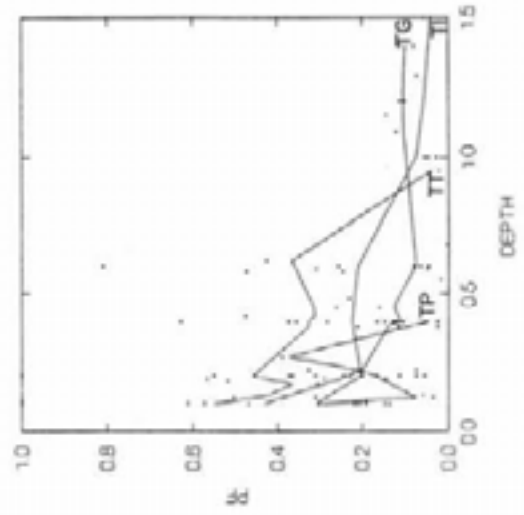
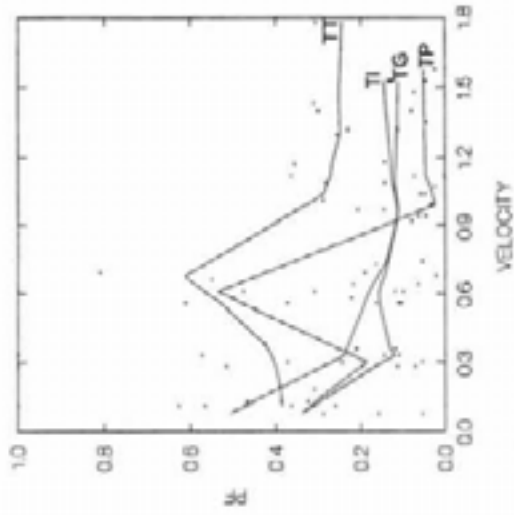
Hydrobiosidae



Chironomidae biomass



Oligochaeta

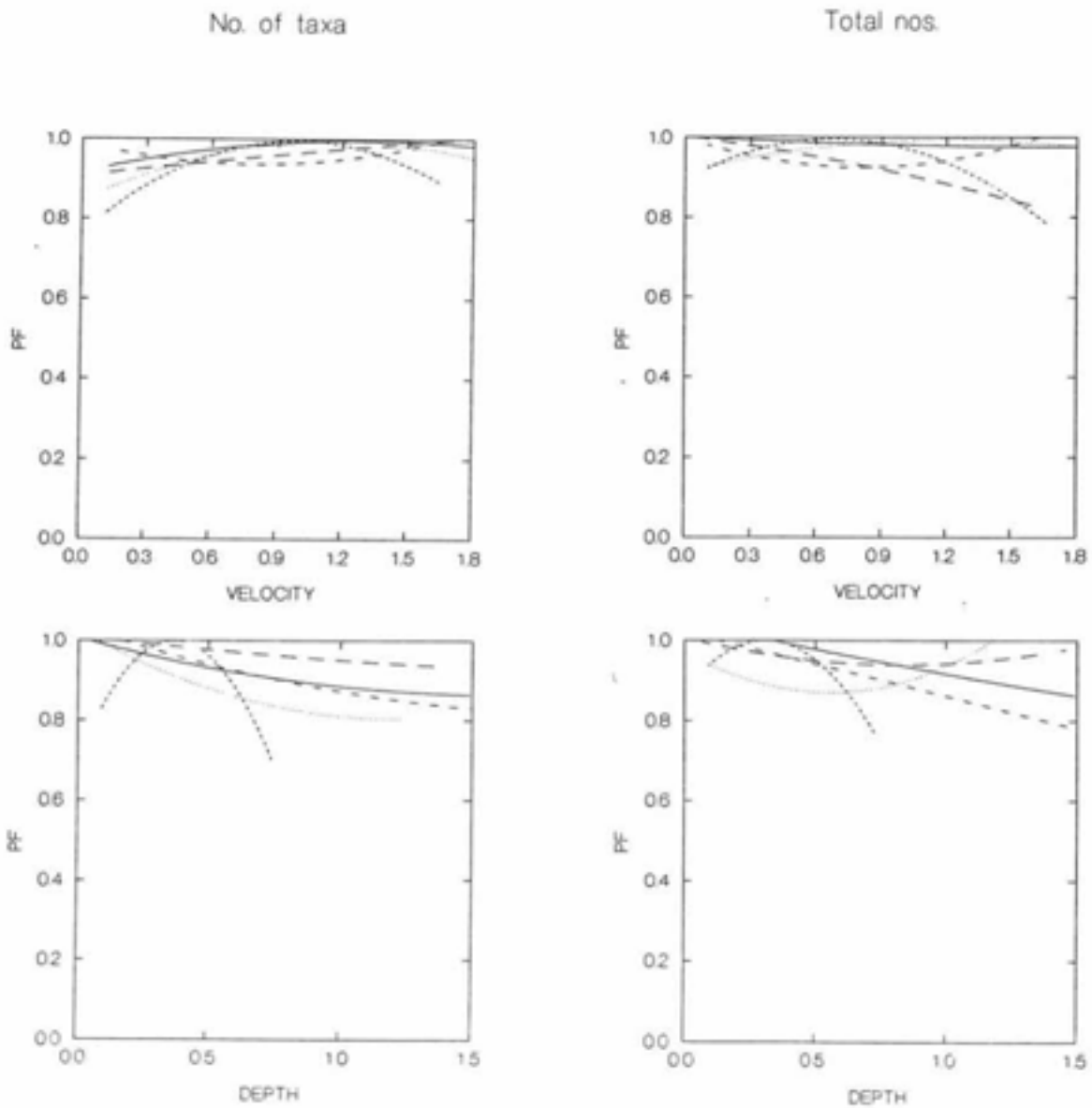




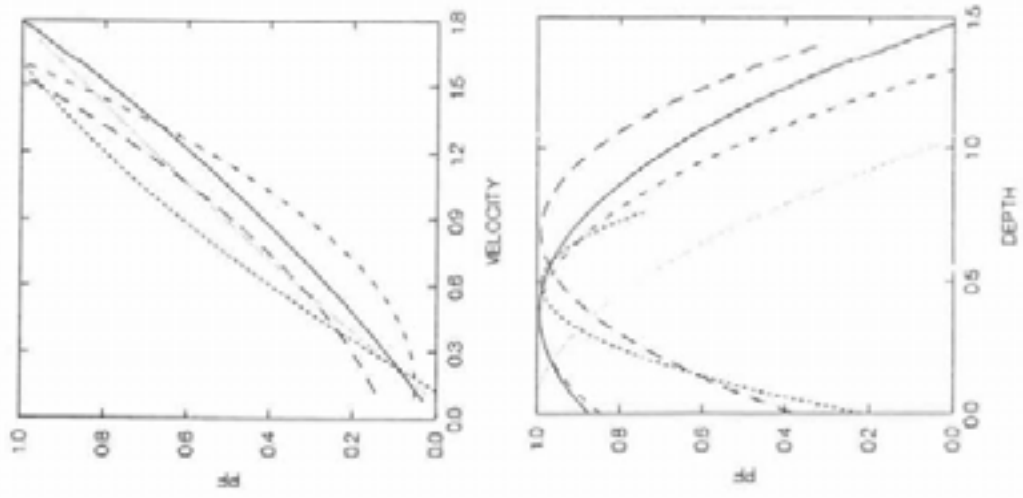
APPENDIX 3B

Exponential polynomial curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for taxonomic richness (number of taxa), total invertebrate densities, densities of 7 major taxa and chironomid biomass in relation to water velocity ( $m.s^{-1}$ ) and depth (m) at 3 sites on Tongariro River.

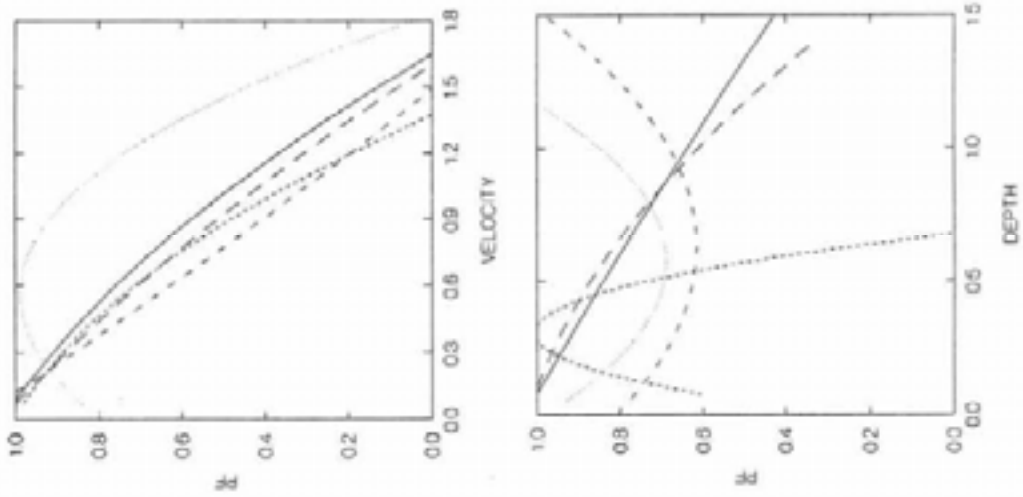
LEGEND: Solid line = All sites combined  
 Long dashes = Site TG  
 Medium dashes = Site TI  
 Short dashes = Site TP  
 Dotted line = Site TT



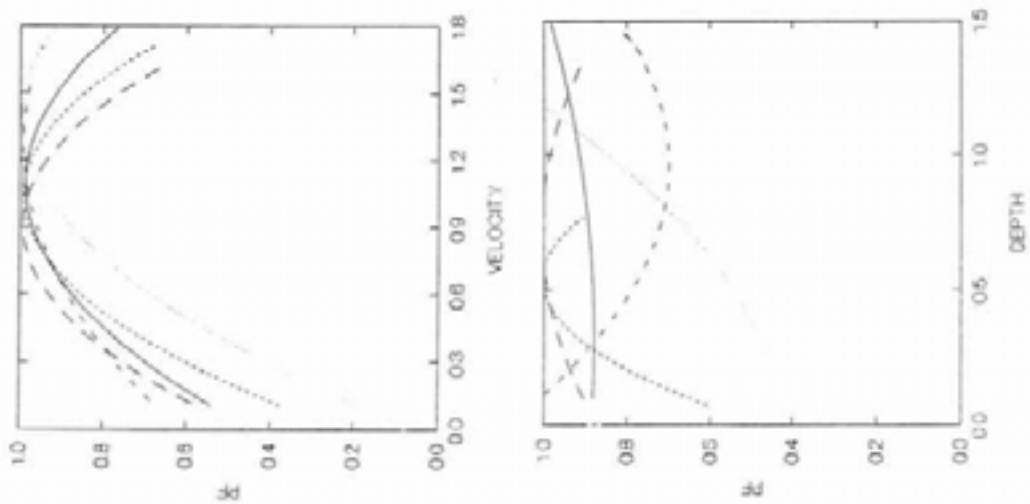
*Aoteapsyche* spp.



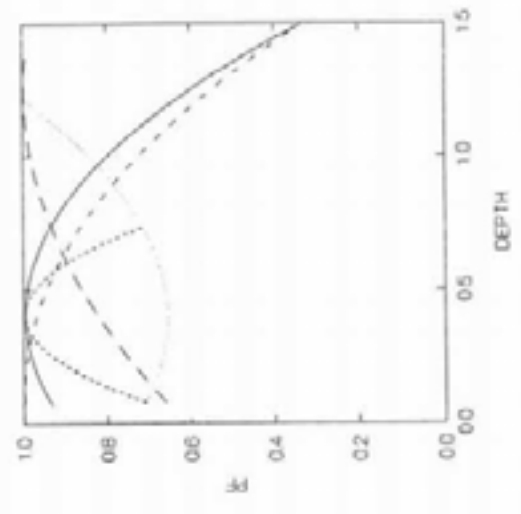
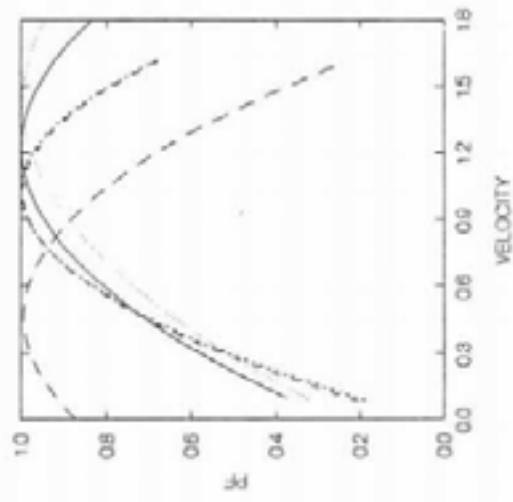
*Z. furcillatus*



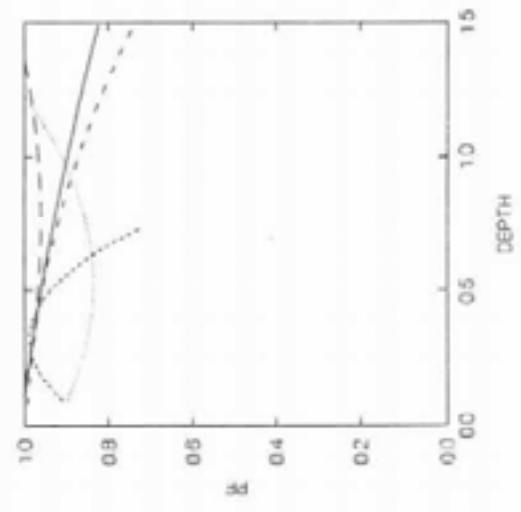
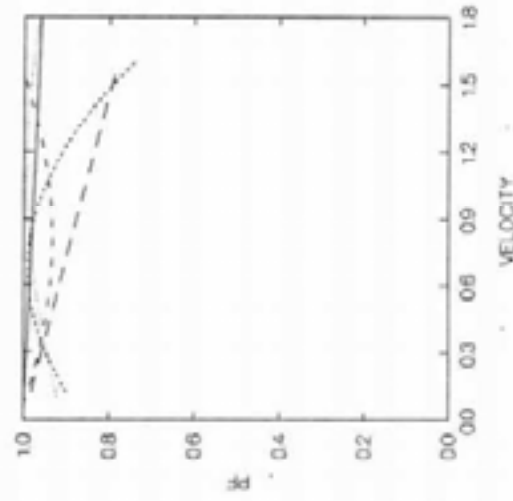
*Deleaticium* spp.



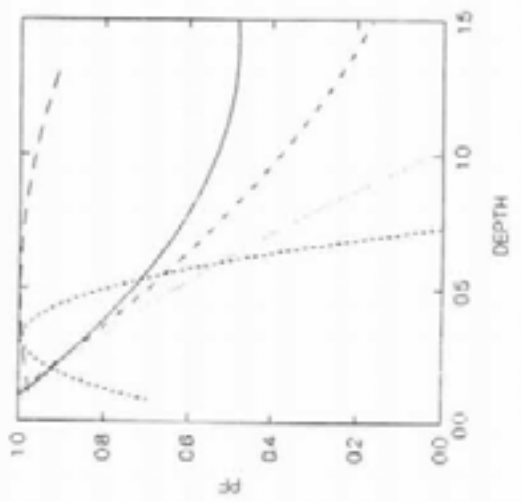
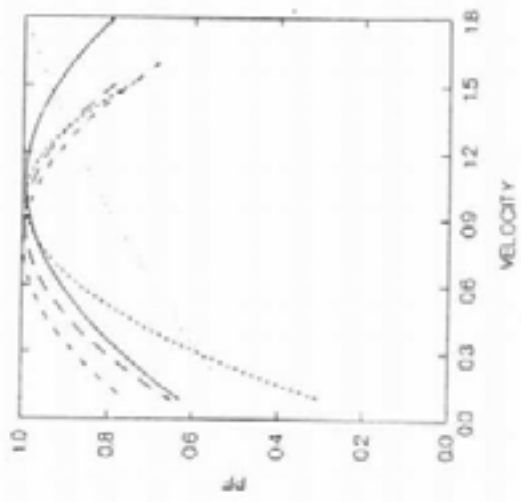
*A. neozelandica*



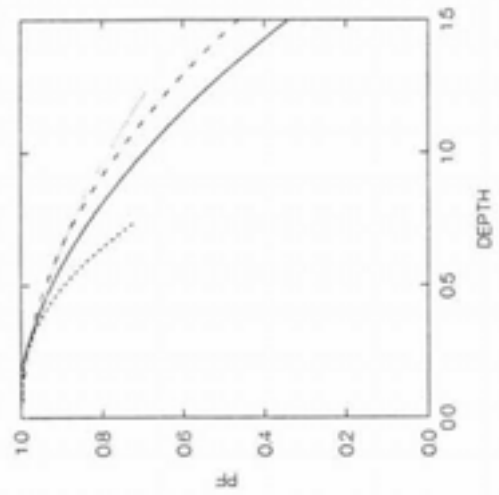
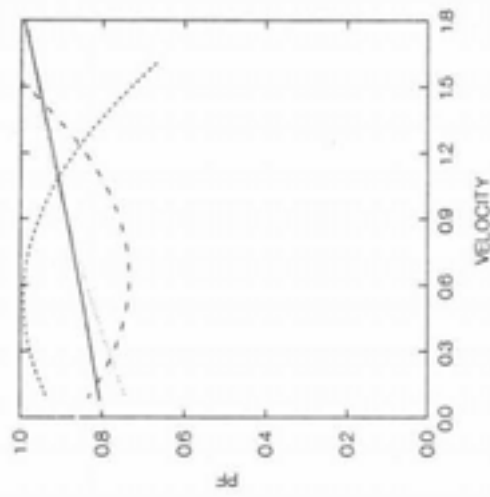
Chironomidae



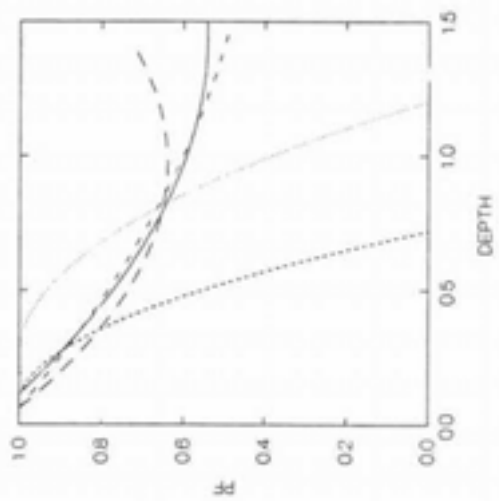
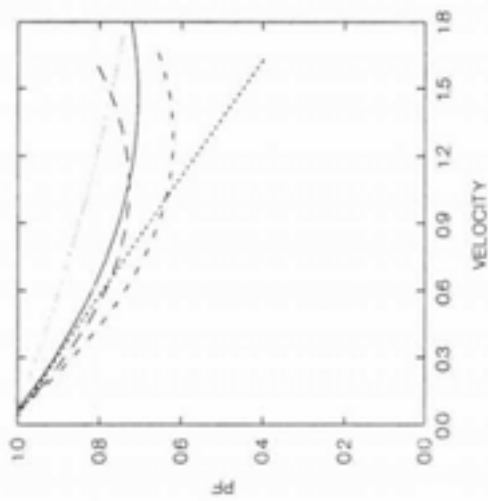
Hydrobiosidae



Chironomidae biomass

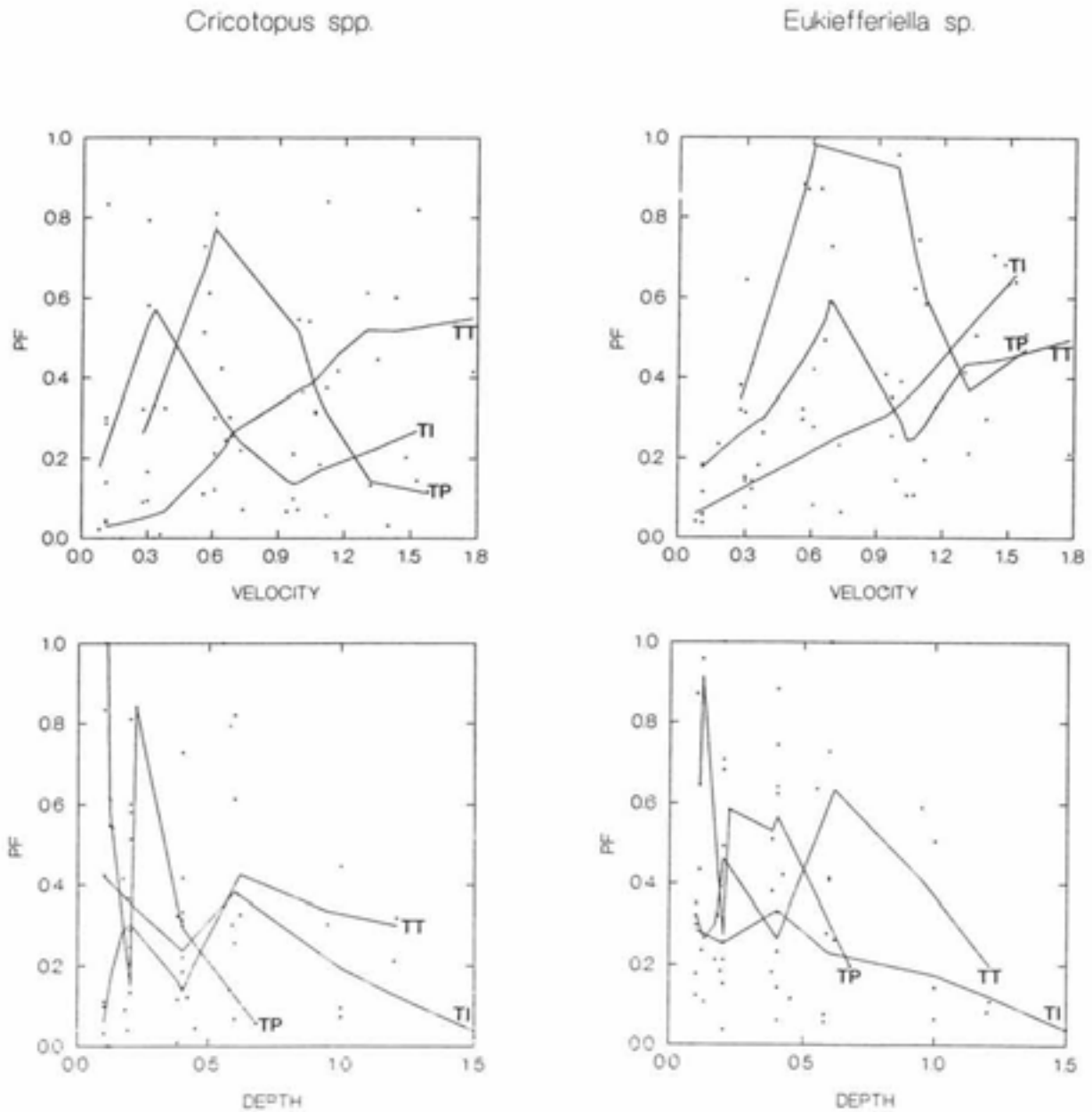


Oligochaeta

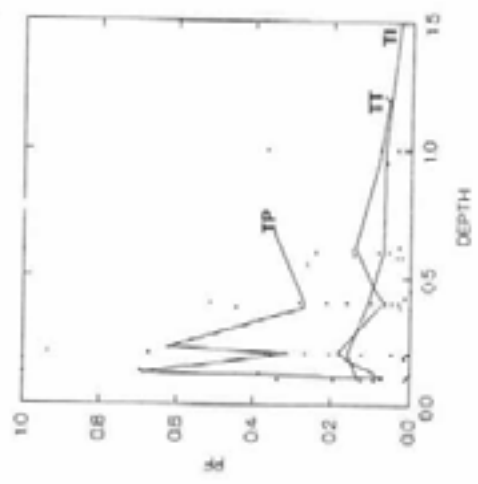
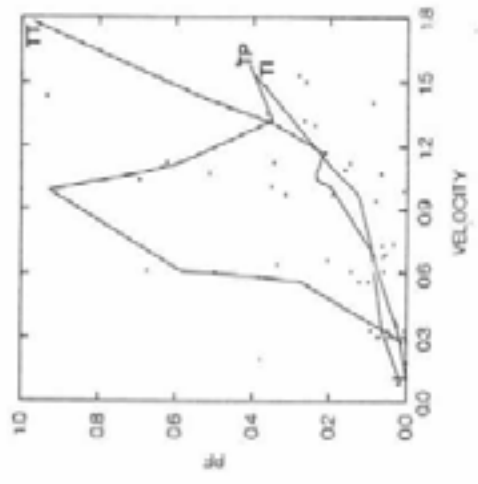


#### APPENDIX 4A

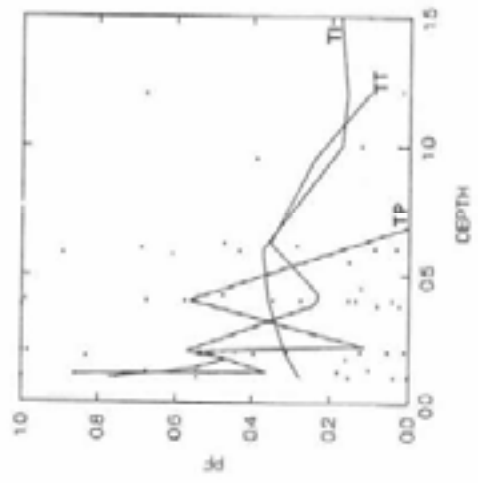
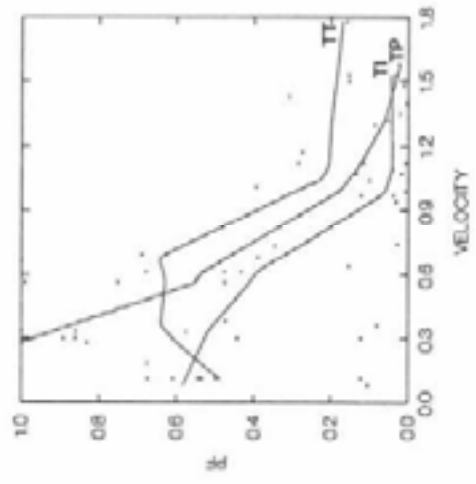
Locally weighted smooth curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for densities of 5 major chironomid taxa in relation to water velocity ( $\text{m.s}^{-1}$ ) and depth (m) at 3 sites on Tongariro River.



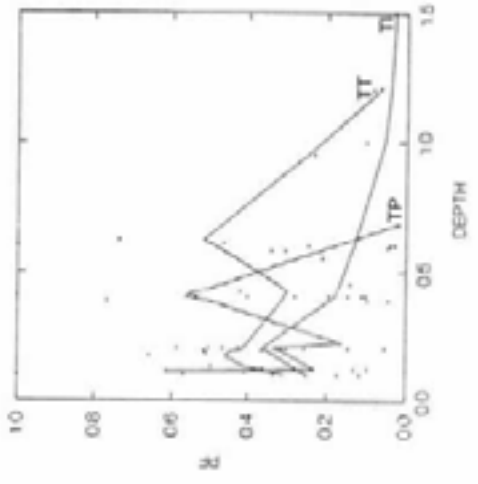
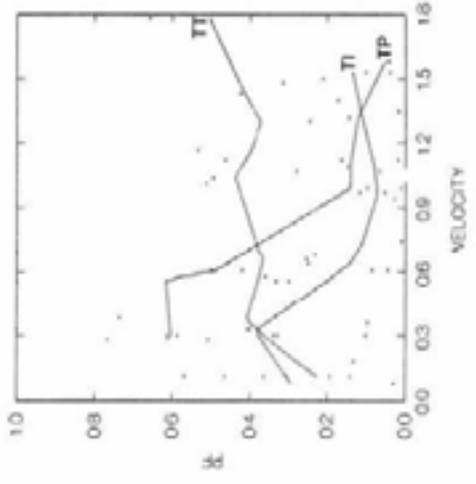
*Macridiamesa* sp.



*Tanytarsus vespertinus*



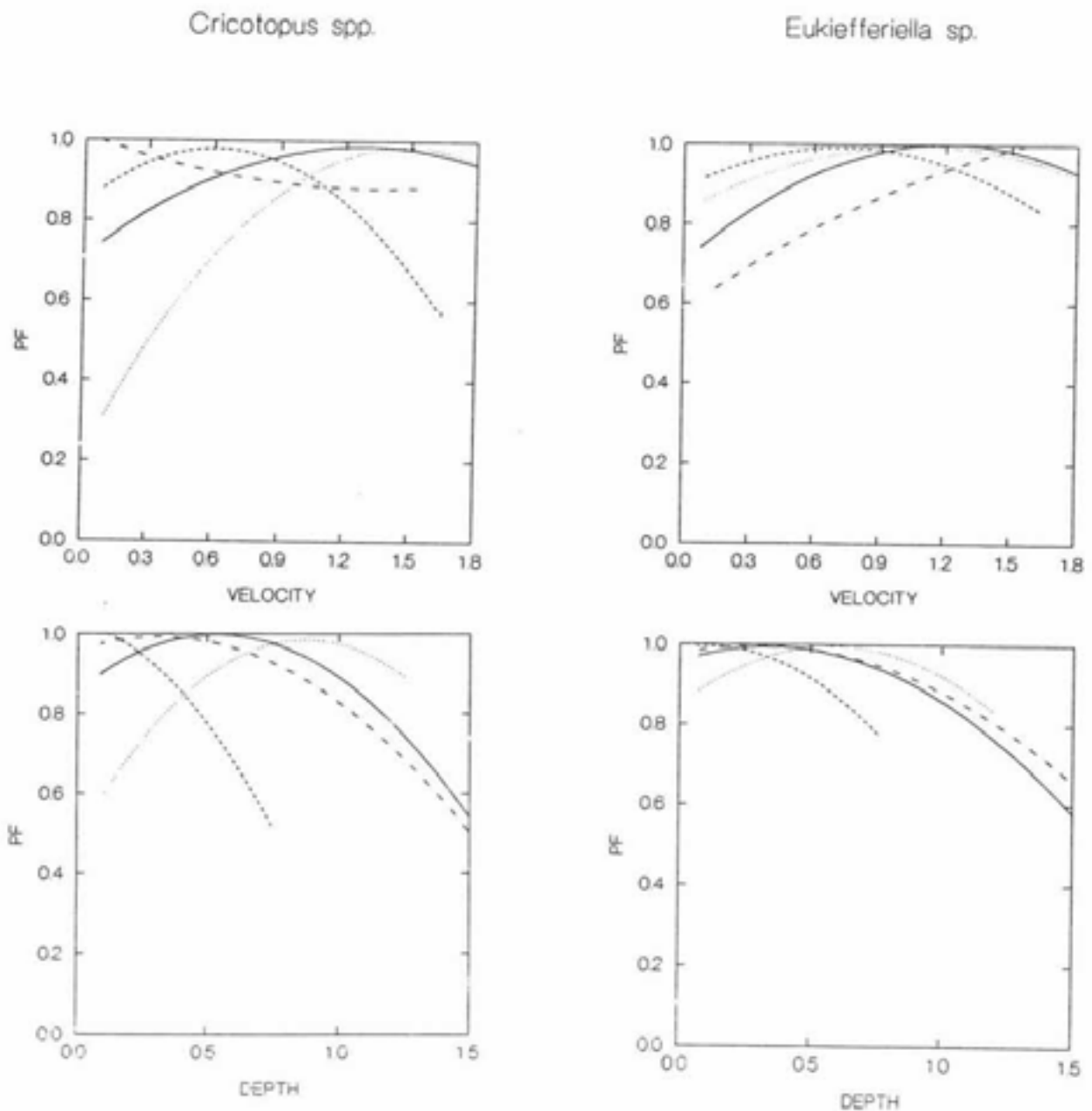
*Orthocleidiinae* sp.a



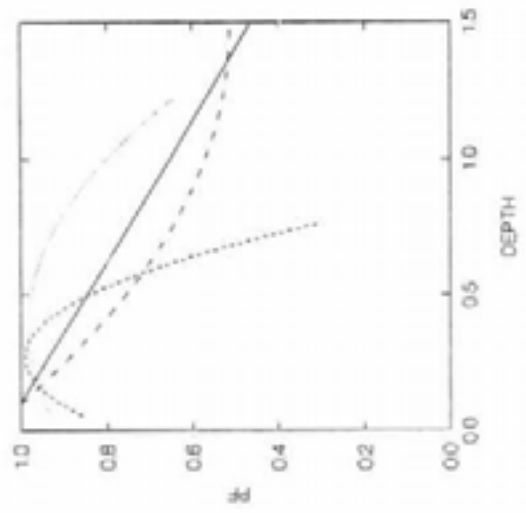
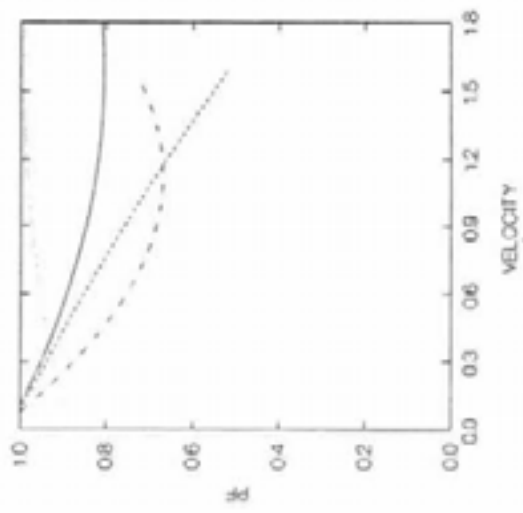
APPENDIX 4B

Exponential polynomial curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for densities of 5 major chironomid taxa in relation to water velocity (m.s-1) and depth (m) at 3 sites on Tongariro River.

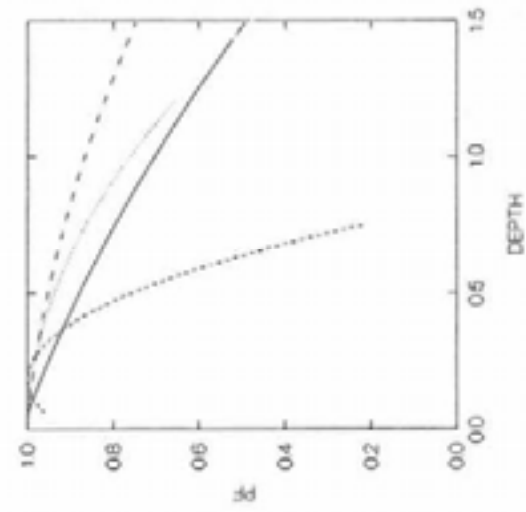
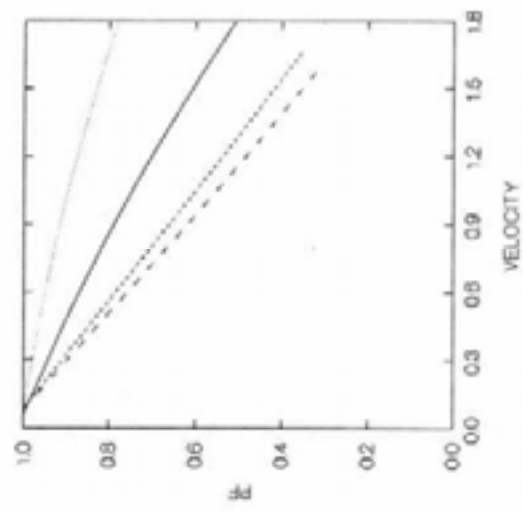
LEGEND: Solid line = All sites combined  
 Medium dashes = Site TI  
 Short dashes = Site TP  
 Dotted line = Site TT



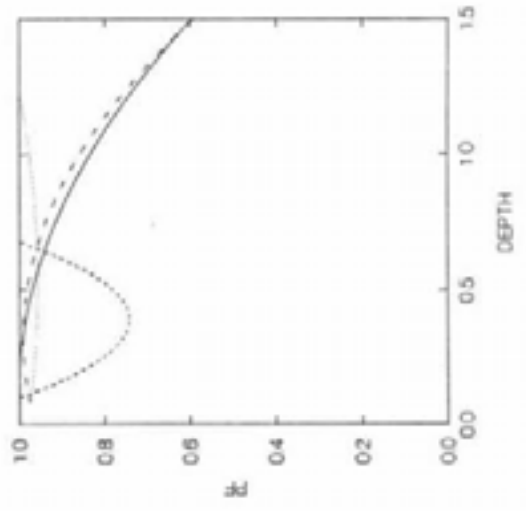
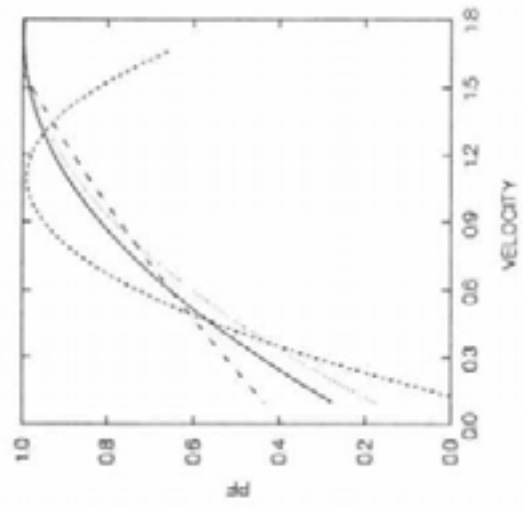
Orthocladinae sp.



Tanytarsus vespertinus



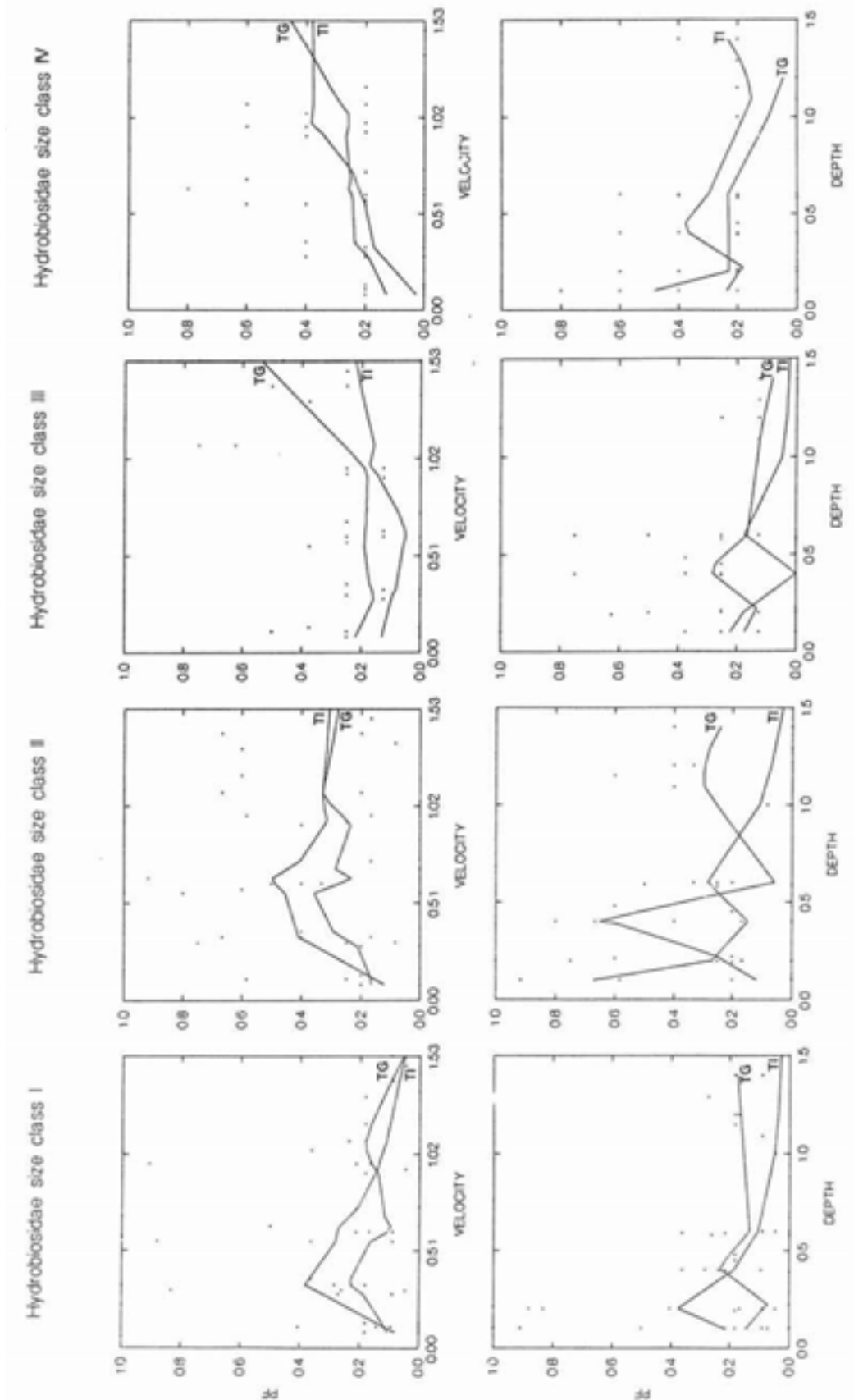
Maoriidamesa sp.





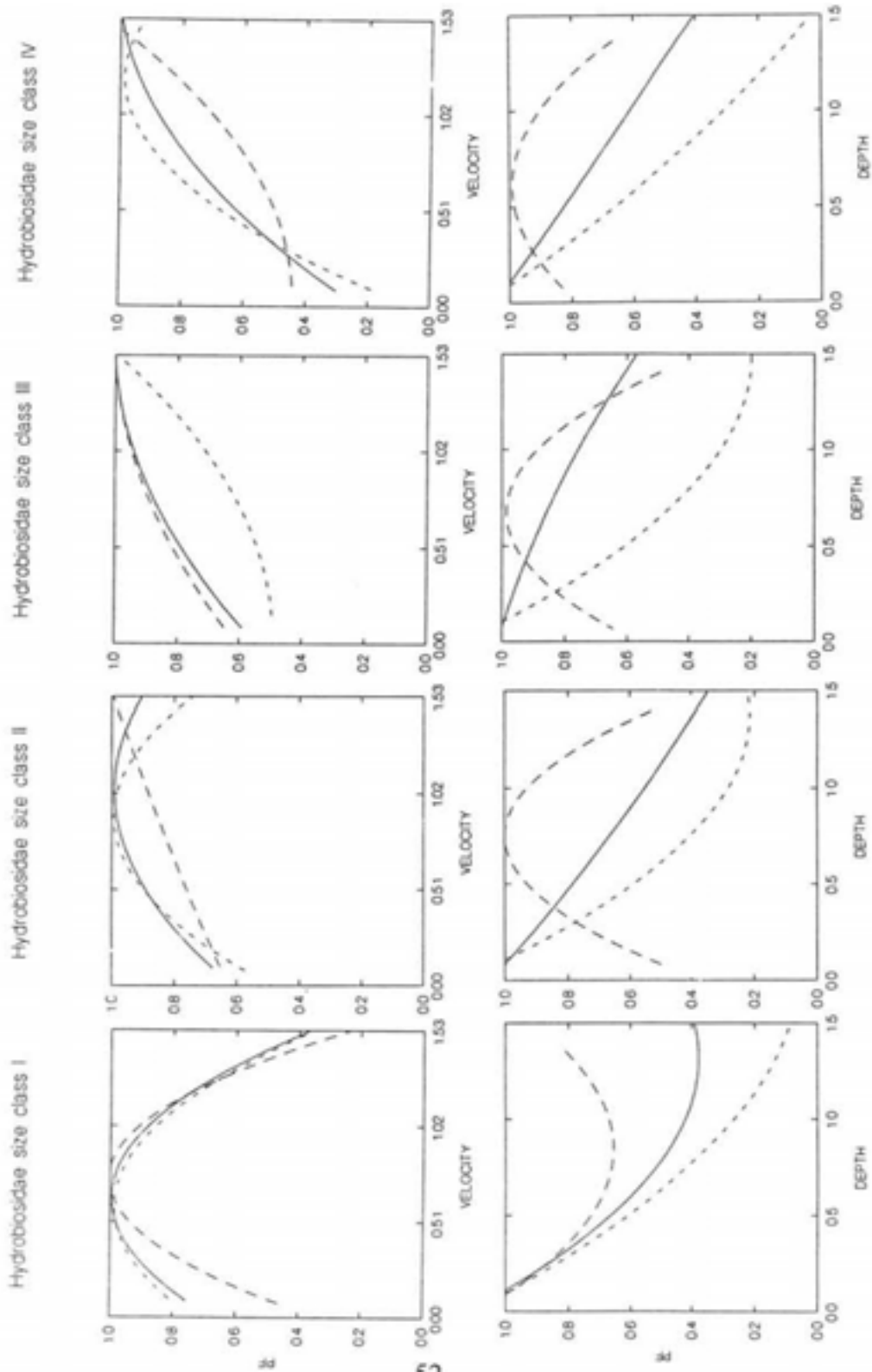
APPENDIX 5A

Locally weighed smooth curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for densities of 4 size classes of Hydrobiosidae larvae in relation to water velocity ( $m.s^{-1}$ ) and depth (m) at 3 sites on Tongariro River.



### APPENDIX 5B

Exponential polynomial curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for densities of 4 size classes of Hydrobiosidae larvae in relation to water velocity ( $\text{m.s}^{-1}$ ) and depth (m) at 3 sites on Tongariro River.

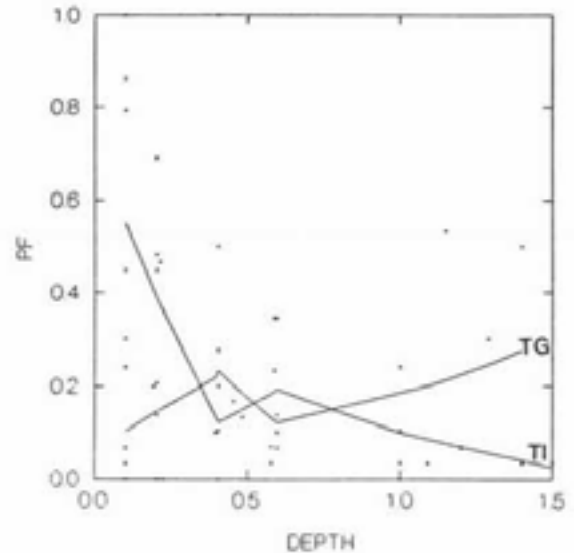
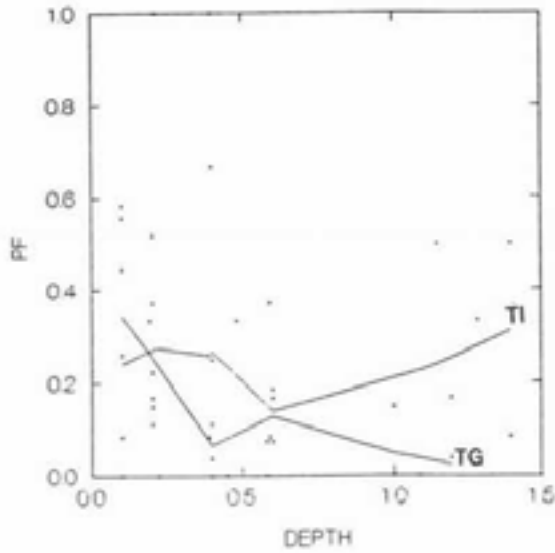
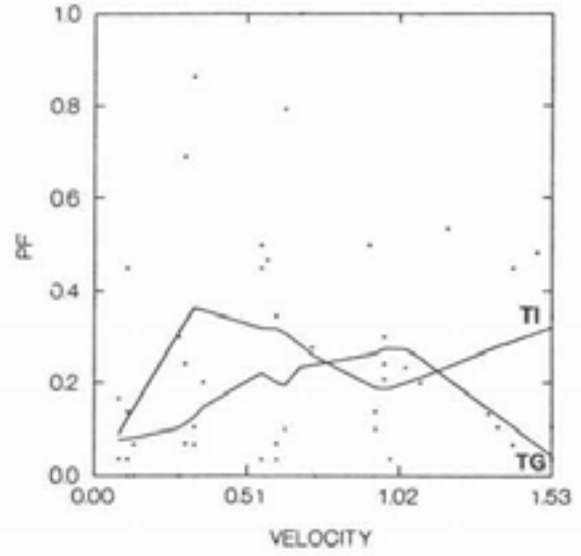
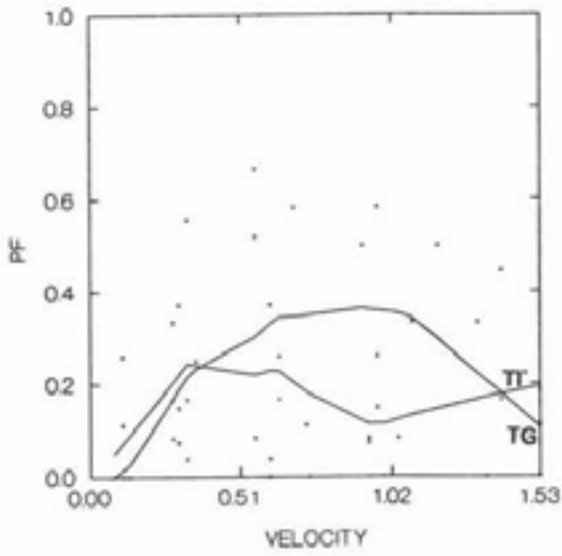


APPENDIX 6A

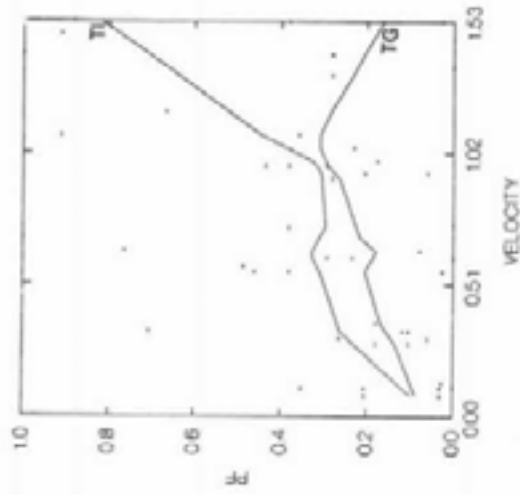
Locally weighted smooth curves plotted through preference factors (PF) (see Section 2.5 for details on calculation) for densities of 8 size classes of *Deleatidium* spp. nymphs in relation to water velocity ( $\text{m.s}^{-1}$ ) and depth (m) at 2 sites on Tongariro River.

Deleatidium size class I

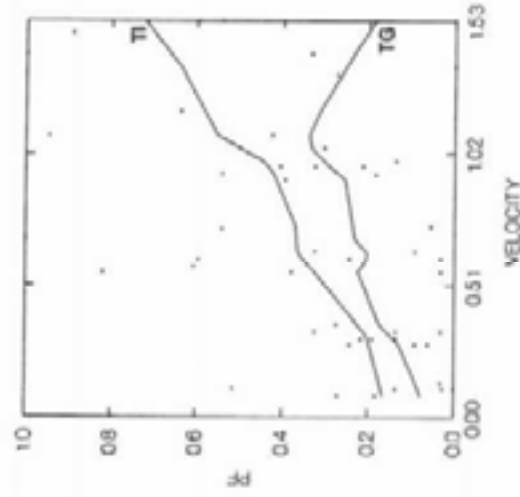
Deleatidium size class II



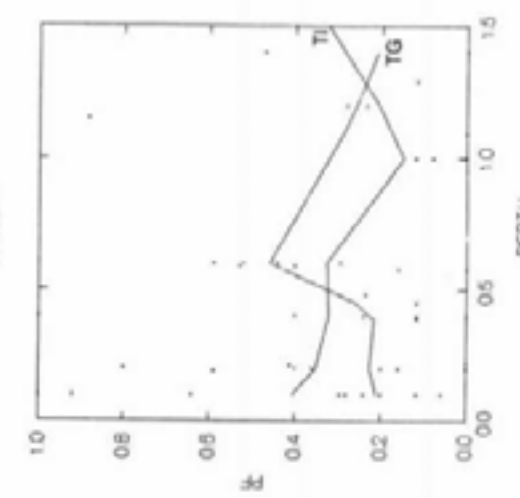
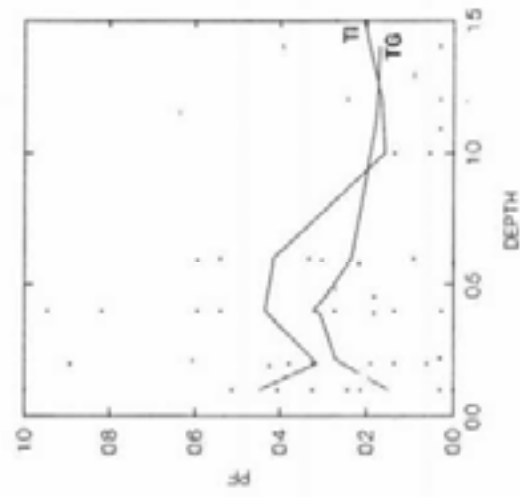
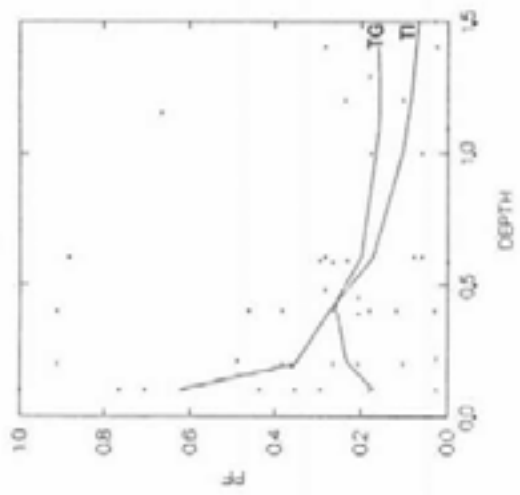
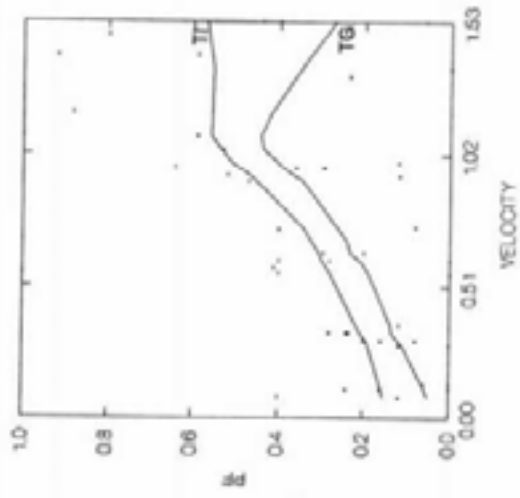
Deleatidium size class III



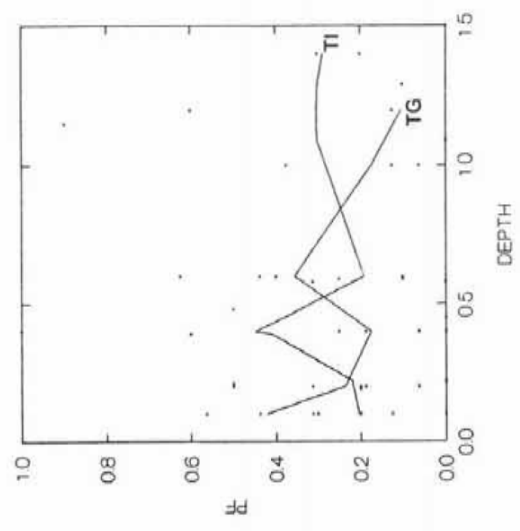
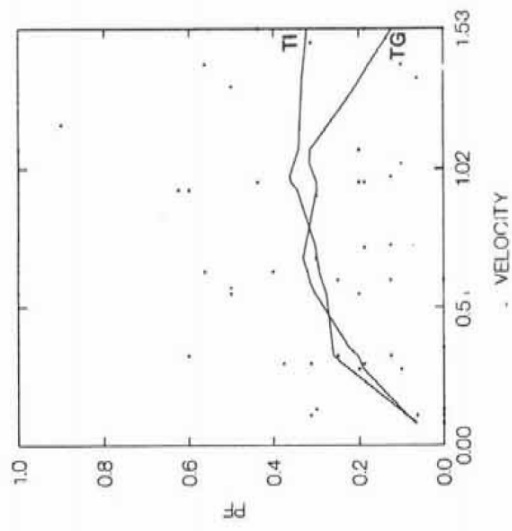
Deleatidium size class IV



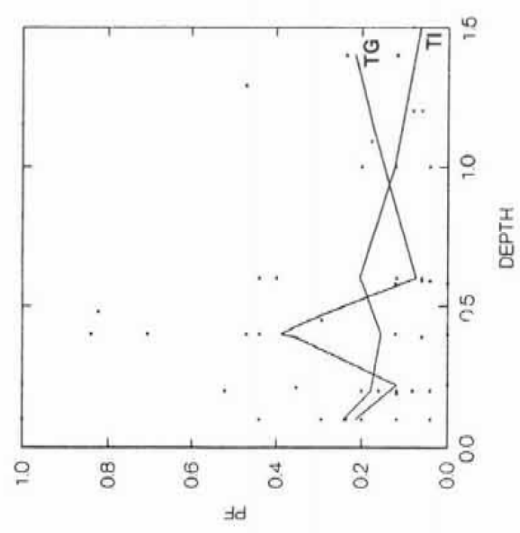
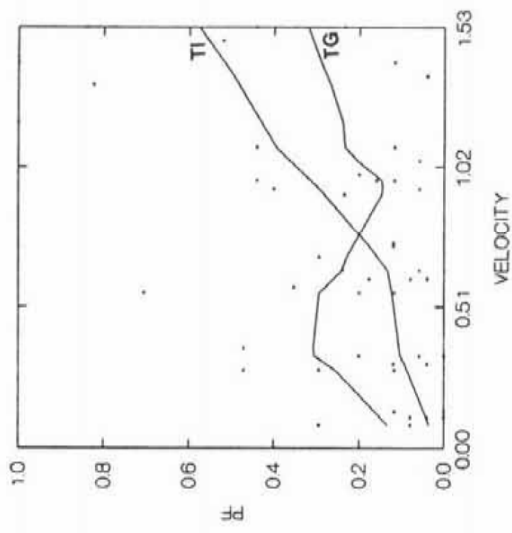
Deleatidium size class V



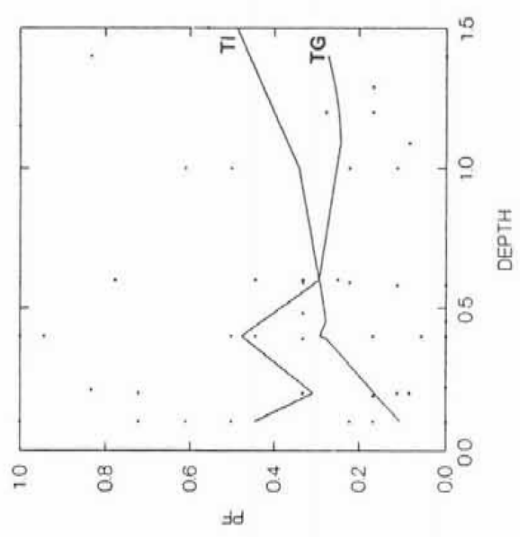
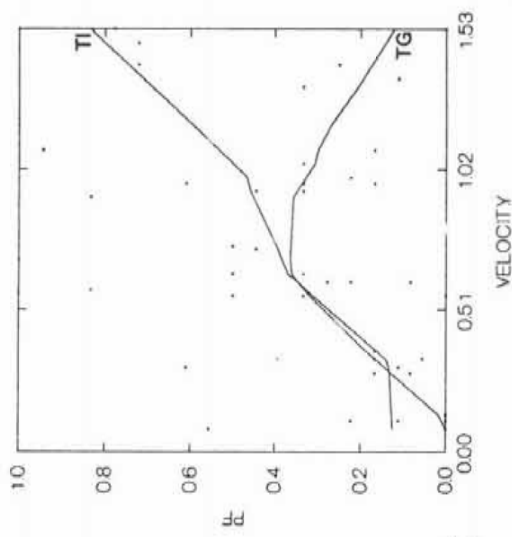
Deleatidium size class VIII



Deleatidium size class VII



Deleatidium size class VI



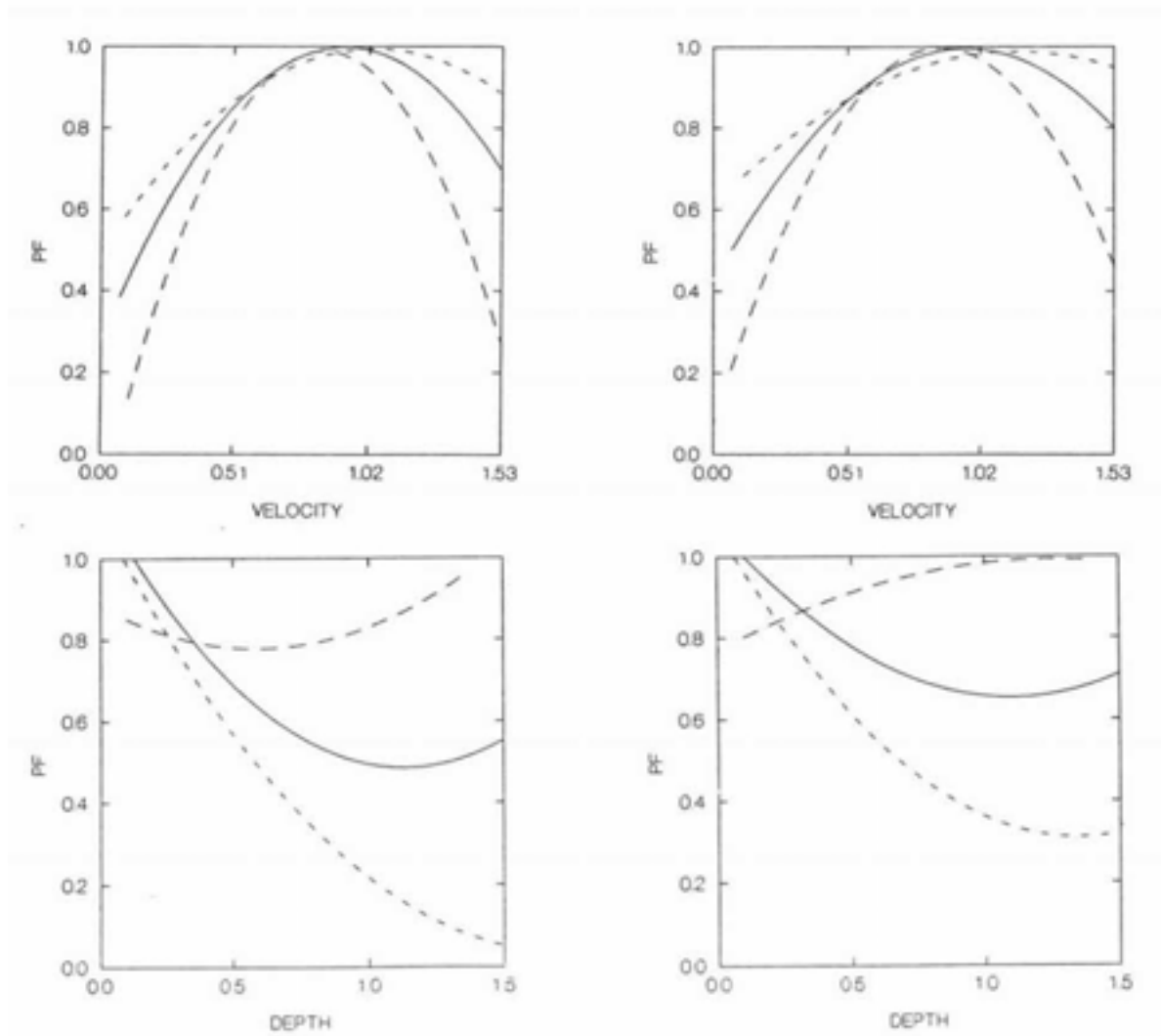
APPENDIX 6B

Exponential polynomial curves plotted through preference factors (PF) (see Section 2.5 for details of calculation) for densities of 8 size classes of *Deleatidium* spp. nymphs in relation to water velocity ( $\text{m.s}^{-1}$ ) and depth (m) at 3 sites on Tongariro River.

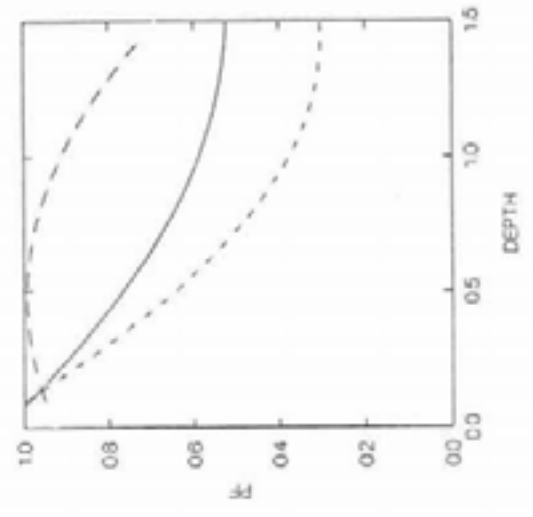
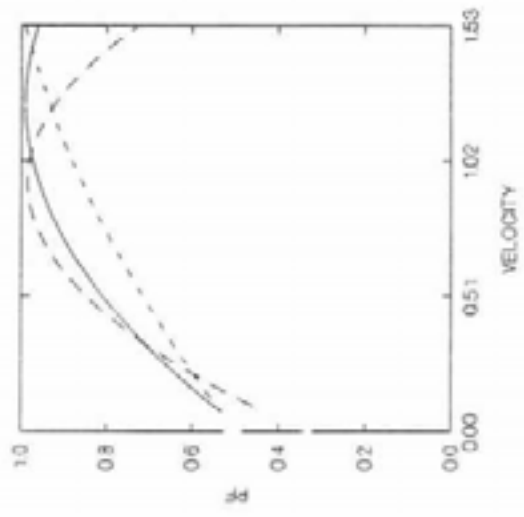
LEGEND: Solid line = all sites combined  
 Long dashes = Site TG  
 Medium dashes = Site TI

Deleatidium size class I

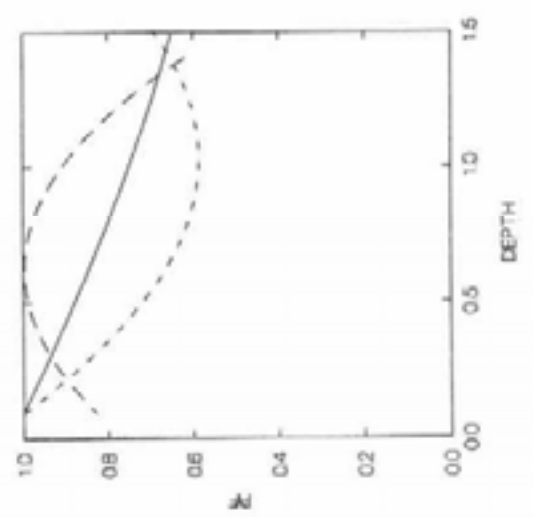
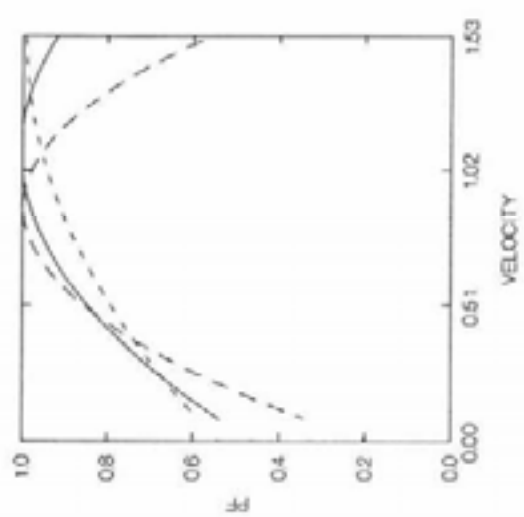
Deleatidium size class II



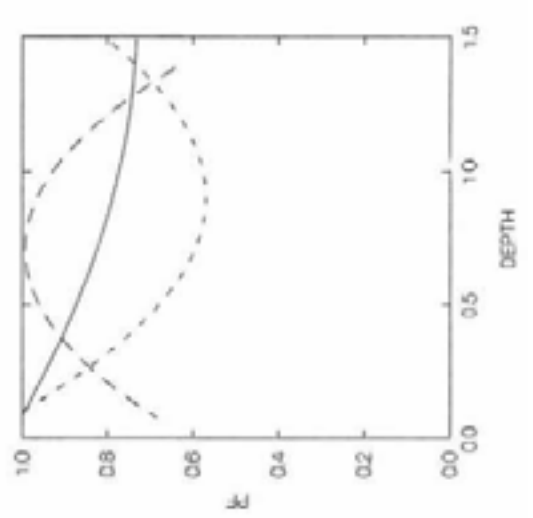
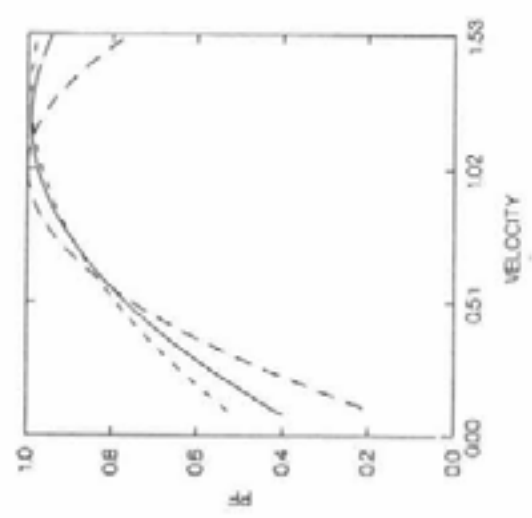
Deleatidium size class III



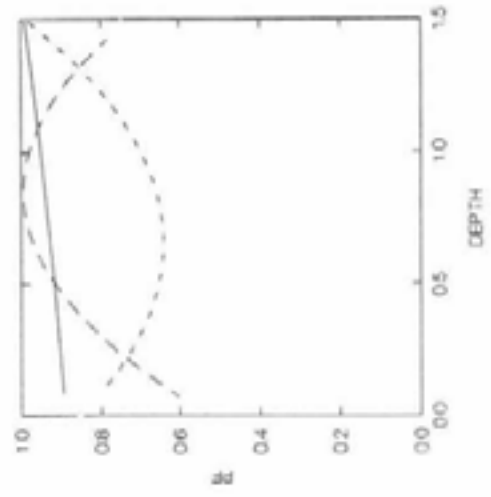
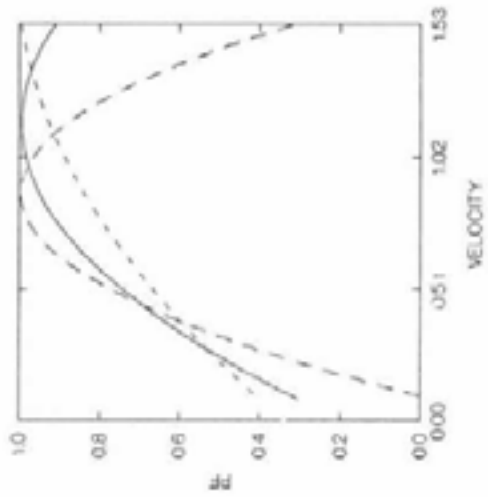
Deleatidium size class IV



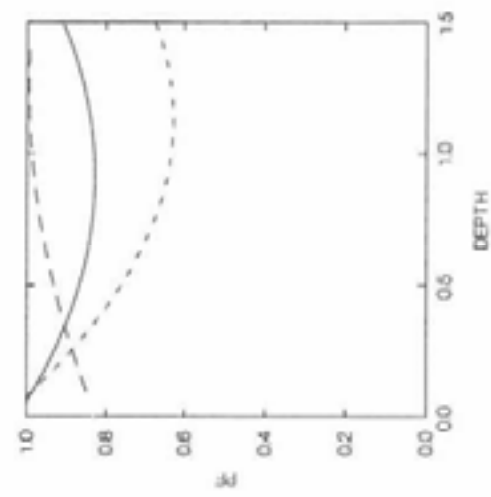
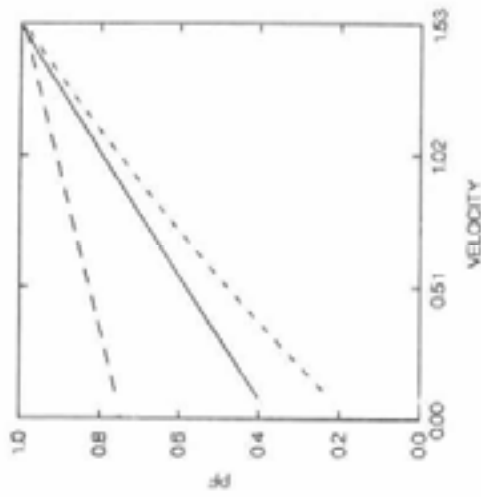
Deleatidium size class V



Deleatidium size class VI



Deleatidium size class VII



Deleatidium size class VIII

